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<p>(54) Title: REPORTER GENE SYSTEM FOR USE IN CELL-BASED ASSESSMENT OF INHIBITORS OF THE HEPATITIS C VIRUS PROTEASE</p> <p><b>(57) Abstract</b></p> <p>A cell-based assay system in which the detection of the reporter gene activity, or secreted alkaline phosphatase (SEAP), is dependent upon the protease activity of the Hepatitis C virus NS3 gene product. This system can be used to assess the activity of candidate protease inhibitors in a mammalian cell-based assay system. The assay system is simpler than previously described assays due to the use of SEAP which allows the reporter gene activity to be quantified by measuring the amount of secreted gene product in the cell media by monitoring the conversion of luminescent or colorimetric alkaline phosphatase substrate.</p>			

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**Reporter Gene System For Use In Cell-Based Assessment  
Of Inhibitors Of The Hepatitis C Virus Protease**

**Technical and Industrial Applicability of Invention**

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A cell-based assay system in which the detection of reporter gene activity (secreted alkaline phosphatase or SEAP) is dependent upon active Hepatitis C virus (HCV) NS3 protease. The assay system is useful in the *in vitro* screening, in a mammalian cell-based assay, of potential protease inhibiting molecules useful in the treatment of HCV. The advantages of using SEAP over more routinely used reporter genes such as beta-galactosidase or luciferase, is that a cell lysis step is not required since the SEAP protein is secreted out of the cell. The absence of a cell lysis step decreases intra- and inter-assay variability as well as makes the assay easier to perform than earlier assays.

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**Background of The Invention**

HCV is one of the major causes of parenterally transmitted non-A, non-B hepatitis worldwide. HCV is now known as the etiologic agent for Non-A Non-B hepatitis throughout the world. Mishiro et al., U.S. Patent No. 5,077,193; Mishiro et al., U.S. Patent No. 5,176,994; Takahashi et al., U.S. Patent No. 5,032,511; Houghton et al., U.S. Patent Nos. 5,714,596 and 5,712,088; as well as (M. Houghton, *Hepatitis C Viruses*, p.1035-1058 in B.N. Fields et al.(eds.), Field's Virology (3d. ed. 1996). HCV infection is characterized by the high rate (>70%) with which acute infection progresses to chronic infection (Alter, M. J. 1995. Epidemiology of hepatitis C in the west. Sem. Liver Dis. 15:5-14.). Chronic HCV infection may lead to progressive liver injury, cirrhosis, and in some cases, hepatocellular carcinoma. Currently, there are no specific antiviral agents available for the treatment of HCV infection. Although alpha interferon therapy is often used in the treatment of HCV-induced moderate or severe liver disease, only a minority of patients exhibit a sustained response Saracco, G. et al., J. Gastroenterol. Hepatol. 10:668-673 1995. Additionally, a vaccine to prevent HCV infection is not yet available and it remains uncertain whether vaccine development will be complicated by the existence of multiple HCV genotypes as well as viral

variation within infected individuals Martell, M. et al., J. Virol. 66:3225-3229 1992; Weiner, et al., Proc. Natl. Acad. Sci. 89:3468-3472 1992. The presence of viral heterogeneity may increase the likelihood that drug resistant virus will emerge in infected individuals unless antiviral therapy effectively suppresses

5 virus replication. Most recently, several of the HCV encoded enzymes, specifically the NS3 protease and NS5B RNA polymerase, have been the focus of intensive research, in vitro screening, and/or rational drug design efforts.

10 HCV has been classified in the flavivirus family in a genus separate from that of the flaviviruses and the pestiviruses. Rice, C. M., in B. N. Fields and P. M. Knipe (eds.), Virology, 3rd edn., p. 931-959; 1996 Lippincott-Raven, Philadelphia, PA. Although the study of HCV replication is limited by the lack of an efficient cell-based replication system, an understanding of replicative

15 events has been inferred from analogies made to the flaviviruses, pestiviruses, and other positive strand RNA viruses. The HCV virus has a 9.4 kb single positive-strand RNA genome encoding over 3,000 amino acids. The genome expresses over 10 structural and non-structural proteins. Post-translational processing of the viral genome requires cleavage by two proteases. As in the

20 pestiviruses, translation of the large open reading frame occurs by a cap-independent mechanism and results in the production of a polyprotein of 3010-3030 amino acids. Proteolytic processing of the structural proteins (the nucleocapsid protein or core (C)) and two envelope glycoproteins, E1 and E2 is accomplished by the action of host cell signal peptidases. Santolini, E., et al., J. Virol. 68:3631-3641, 1994; Ralston, R., et al., J. Virol. 67:6753-6761

25 1993. Cleavage of the nonstructural proteins (NS4A, NS4B, NS5A, and NS5B) is mediated by the action of the NS2/3 protease or the NS3 protease. Grakoui, A. et al., J. Virol. 67:2832-2843 1993; Hirowatari, Y., et al., Anal. Biochem. 225:113-120 1995; Bartenschlager, R. et al., J. Virol. 68:5045-5055

30 1994; Eckart, M. R., et al., Biochem. Biophys. Res. Comm. 192:399-406 1993; Grakoui, A., et al., J. Virol. 67:2832-2843 1993; Tomei, L., et al., J. Virol. 67:4017-4026 1993; NS4A is a cofactor for NS3 and NS5B is an RNA dependent RNA polymerase. Bartenschlager, R. et al., (1994); Failla, C., et al., J. Virol. 68:3753-3760 1994; Lin, C. et al., Proc. Natl. Acad. Sci. 92:7622-

7626 1995; Behrens, S.-E., et al., EMBO J. 15:12-22 1996. Functions for the NS4B and NS5A proteins have yet to be defined.

The NS2/3 is a metalloprotease and has been shown to mediate cleavage at  
5 the 2/3 junction site Grakoui, et al. (1993); Hijikata, M., et al., J. Virol. 67:4665-4675  
1993. In contrast, the NS3 protease is required for multiple cleavages within the  
nonstructural segment of the polyprotein, specifically the 3/4A, 4A/4B, 4B/5A, and  
5A/5B junction sites Bartenschlager et al. (1993); Eckart, M. R., et al., Biochem.  
Biophys. Res. Comm. 192:399-406 1993; Grakoui et al. (1993); Tomei et al. (1994).  
10 More recently, it is thought that the NS2/3 protease might actually be part of the HCV  
NS3 protease complex even though they have two functionally distinct activities.  
Although NS3 protease is presumed to be essential for HCV viability, definitive proof  
of its necessity has been hampered by the lack of an infectious molecular clone that  
can be used in cell-based experiments. However, recently two independent HCV  
15 infectious molecular clones have been developed and have been shown to replicate  
in chimpanzees. Kolykhalov, A. A., et al., Science 277:570-574 1997; Yanagi, M., et  
al., Proc. Natl. Acad. Sci. 94:8738-8743 1997. The requirement for NS3 in the HCV  
life cycle may be validated in these clones by using oligo nucleotide-mediated site  
directed mutagenesis to inactivate the NS3 catalytic serine residue and then  
20 determining whether infectious virus is produced in chimpanzees. Until these  
experiments are performed, the necessity of NS3 is inferred from cell-based  
experiments using the related yellow fever (YFV) and bovine viral diarrhea (BVDV)  
viruses. Mutagenesis of the YFV and BVDV NS3 protease homologs has shown that  
NS3 serine protease activity is essential for YFV and BVDV replication. Chambers, T.  
25 J., et al., Proc. Natl. Acad. Sci. 87:8898-8902 1990; Xu, J., et al., J. Virol. 71:5312-  
5322 1997.

In general, when investigators screen potential anti-viral compounds for  
inhibitory activity, it usually involves initial *in vitro* testing of putative enzyme inhibitors  
30 followed by testing the compounds on actual infected cell lines and animals. It is  
obvious that working with live virus in large scale screening activities can be  
inherently dangerous and problematic. While final testing of putative inhibitors in  
infected cells and animals is still necessary for preclinical drug development, for initial  
screening of candidate molecules, such work is cost-prohibitive and unnecessary.  
35 Furthermore, the inability to grow HCV in tissue culture in a reproducible quantitative

manner prevents the evaluation of potential antiviral agents for HCV in a standard antiviral cytopathic effect assay. In response to this real need in the industry, development of non-infectious, cell-based, screening systems is essential.

5       For example, Hirowatari, et al. developed a reporter assay system, *inter alia*, that involves the transfection of mammalian cells with two eukaryotic expression plasmids. Hirowatari, et al., *Anal. Biochem.* 225:113-120 1995. One plasmid has been constructed to express a polyprotein that encompasses the HCV NS2-NS3 domains fused in frame to an NS3 cleavage site followed by the HTLV-1 TAX1

10      protein. A second plasmid has been constructed to have the expression of the chloramphenicol acetyltransferase (CAT) reporter gene under the control of the HTLV-1 LTR. Thus when COS cells are transfected with both plasmids, NS3-mediated cleavage of the TAX1 protein from the NS2-NS3-TAX1 polyprotein allows the translocation of TAX1 to the nucleus and subsequent activation of CAT

15      transcription from the HTLV-1 LTR. CAT activity can be measured by assaying the acetylation of <sup>14</sup>C-chloramphenicol through chromatographic or immunological methods. In the CAT assay generally, cell extracts are incubated in a reaction mix containing <sup>14</sup>C- or <sup>3</sup>H-labeled chloramphenicol and n-Butyryl Coenzyme A. The CAT enzyme transfers the n-butyryl moiety of the cofactor to chloramphenicol. For a

20      radiometric scintillation detection (LSC) assay, the reaction products are extracted with a small volume of xylene. The n-butyryl chloramphenicol partitions mainly into the xylene phase, while unmodified chloramphenicol remains predominantly in the aqueous phase. The xylene phase is mixed with a liquid scintillant and counted in a scintillation counter. The assay can be completed in as little as 2-3 hours, is linear for

25      nearly three orders of magnitude, and can detect as little as  $3 \times 10^4$  units of CAT activity. CAT activity also can be analyzed using thin layer chromatography (TLC). This method is more time-consuming than the LSC assay, but allows visual confirmation of the data.

30      Similarly, the other patents of Houghton, et al., U.S. Patent No. 5,371,017, U.S. Patent No. 5,585,258, U.S. Patent No. 5,679,342 and U.S. Patent No. 5,597,691 or Jang et al. WO 98/00548 all disclose a cloned NS3 protease or portion fused to a second gene encoding for a protein which a surrogate expression product can be detected for example, in the '017 patent of Houghton, b-galactosidase, superoxide dismutase, ubiquitin or in Jang, the expression is measured by the proliferation of

poliovirus in cell culture) and its use for candidate screening. It is unclear in the Houghton, et al. patents, however, whether the protease described in the specification is the NS2/3 metalloprotease or NS3 serine protease. Although the serine protease is claimed, the experimental data show putative cleavage of the N-terminal SOD fusion partner at the NS2/3 junction, a function which recently has been deemed to be the domain of the NS2/3 metalloprotease (Rice, C.M., et al., Proc. Natl. Acad. Sci. 90:10583-10587 (1993)). Furthermore, an active soluble NS3 serine protease is not disclosed in the Houghton, et al. patents, but a insoluble protein derived from *E. coli* inclusion bodies and which was N-terminally sequenced. For purposes of the present invention the term "NS2 protease" will refer to the enzymatic activity associated with the NS2/3 metalloprotease as defined by Rice et al., and the term "NS3 protease" will refer to the serine protease located within the NS3 region of the HCV genome.

De Francesco et al., U.S. Patent No. 5,739,002, also describes a cell free in vitro system for testing candidates which activate or inhibit NS3 protease by measuring the amount of cleaved substrate. Hirowatari et al. (1995) discloses another HCV NS3 protease assay, however, it differs from the present invention in several aspects, including the reporter gene, the expression plasmid constructs, and the method of detection. Recently, Cho et al. describe a similar SEAP reporter system for assaying HCV NS3 protease which also differs in its structure and function from the present invention. Cho et al., J. Virol. Meth. 72:109-115 1998. Also of interest is a NS3 protease assay system developed by Chen et al. in WO 98/37180. In the Chen et al. application, a fusion protein is described which uses NS3 protease polypeptide or various truncation analogs fused to the NS4A polypeptide or various truncation analogs and is not autocleavable. The fusion protein is then incubated with known substrates with or without inhibitors to screen for inhibitory effect.

There are a number of problems inherent in all the abovementioned assay systems. For example, the reporter gene product or analyte is many steps removed from the initial NS3 protease cleavage step, the cells used in the assay system are prokaryotic or Yeast based and must be lysed before the reporter gene product can be measured, and the surrogate marker is proliferation of live virus. All of these problems are overcome in the present invention as summarized below.

### Summary of Invention

The present invention describes a reporter gene system for use in the cell based assessment of inhibitors of the HCV protease. Applicants point out that 5 throughout the description of this invention, the reference to specific non-structural (NS) regions or domains of the HCV genome are functional definitions and correspond approximately to the defined sequence locations described by C.M. Rice and others. The present invention discloses the co-transfection of a target cell line with a viral vector which has been engineered to express from the T7 RNA 10 polymerase promoter and a recombinant plasmid or viral vector which has been engineered to express a polyprotein that includes NS3 HCV serine protease and the secreted human placental alkaline phosphatase (SEAP) gene (Berger et al. 1988) under control of the T7 promoter. The present invention was designed to have a linkage between the detection of reporter gene activity and NS3 serine protease 15 activity through construction of a segment of the HCV gene encoding the NS2-NS3-NS4A-NS4B'-sequence linked to the SEAP reporter.

Detection of NS3 protease activity is accomplished by having the release and hence, the subsequent detection, of the SEAP reporter gene to be dependent upon 20 NS3 serine protease activity. In a preferred embodiment, the target cell line is first infected with a viral vector that expresses the T7 RNA polymerase followed by either co-infection with a second viral vector that encodes the NS3 HCV protease/SEAP polyprotein, or transfection with a plasmid that contains the same NS3/SEAP gene elements.

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The SEAP enzyme is a truncated form of human placental alkaline phosphatase, in which the cleavage of the transmembrane domain of the protein allows it to be secreted from the cells into the surrounding media. SEAP activity can be detected by a variety of methods including, but not limited to, measurement of 30 catalysis of a fluorescent substrate, immunoprecipitation, HPLC, and radiometric detection. The luminescent method is preferred due to its increased sensitivity over colorimetric detection methods, and such an assay kit is available from Tropix®. The advantages of using SEAP over more routinely used reporter genes such as beta-galactosidase or luciferase, is that a cell lysis step is not required since the SEAP 35 protein is secreted out of the cell. The absence of a cell lysis step decreases intra-

and inter-assay variability as well as makes the assay easier to perform than earlier assays in the prior art. When both the T7 promoter and NS3/SEAP constructs are present, SEAP can be detected in the cell medium within the usual viral assay timeframe of 24-48 hours, however, the timeframe should not be read as a limitation  
5 because it is theoretically possible to detect the SEAP in the media only a few hours after transfection. The medium can then be collected and analyzed. Various examples illustrating the use of this composition and method will be detailed below.

### Brief Description of the Drawings

10                  Figure 1 illustrates schematically the Vaccinia Virus NS3/SEAP System gene construct.  
Figure 1B illustrates schematically the Plasmid/Vaccinia Virus NS3/SEAP assay.  
15                  Figure 2 illustrates schematically how the assay operates.  
Figure 3 illustrates schematically the DI/DR Assay.  
Figure 4A and 4B shows the SEAP activity dose response curve for a representative plasmid/virus assay.  
Figure 5 shows an experimental 96 well plate diagram for the SEAP protocol  
20                  on Day 1 in Example 3.  
Figure 6 shows an experimental 96 well plate diagram for the SEAP protocol on Day 2 in Example 3.  
Figure 7 shows SEAP activity and Cytotoxicity data for Example 4.  
Figure 8 shows a summary of DI/DR assay data.  
25                  Figure 9 illustrates the experimental plate set-up for Example 2.

### Detailed Description of a Preferred Embodiment of the Invention

The practice of this invention will employ, unless otherwise indicated,  
30 conventional techniques of molecular biology, microbiology, recombinant DNA manipulation and production, virology and immunology, which are within the skill of the art. Such techniques are explained fully in the literature: Sambrook, *Molecular Cloning; A Laboratory Manual*, Second Edition (1989); *DNA Cloning*, Volumes I and II (D. N. Glover, Ed. 1985); *Oligonucleotide Synthesis* (M. J. Gait, Ed. 1984); *Nucleic Acid Hybridization* (B. D. Hames and S. I. Higgins, Eds. 1984); *Transcription and*  
35

*Translation* (B. D. Hames and S. I. Higgins, Eds. 1984); *Animal Cell Culture* (R. I. Freshney, Ed. 1986); *Immobilized Cells and Enzymes* (IRL Press, 1986); B. Perbal, *A Practical Guide to Molecular Cloning* (1984); *Gene Transfer Vectors for Mammalian Cells* (J. H. Miller and M. P. Calos, Eds. 1987, Cold Spring Harbor Laboratory);

5     *Methods in Enzymology*, Volumes 154 and 155 (Wu and Grossman, and Wu, Eds., respectively), (Mayer and Walker, Eds.) (1987); *Immunochemical Methods in Cell and Molecular Biology* (Academic Press, London), Scopes, (1987), *Expression of Proteins in Mammalian Cells Using Vaccinia Viral Vectors* in *Current Protocols in Molecular Biology*, Volume 2 (Frederick M. Ausubel, et al., Eds.)(1991). All patents, patent

10    applications and publications mentioned herein, both supra and infra, are hereby incorporated by reference.

Both prokaryotic and eukaryotic host cells are useful for expressing desired coding sequences when appropriate control sequences compatible with the designated host are used. Among prokaryotic hosts, *E. coli* is most frequently used. Expression control sequences for prokaryotes include promoters, optionally containing operator portions, and ribosome binding sites. Transfer vectors compatible with prokaryotic hosts are commonly derived from, for example, pBR322, a plasmid containing operons conferring ampicillin and tetracycline resistance, and the various pUC vectors, which also contain sequences conferring antibiotic resistance markers.

15    These plasmids are commercially available. The markers may be used to obtain successful transformants by selection. Commonly used prokaryotic control sequences include the  $\beta$ -lactamase (penicillinase) and lactose promoter systems (Chang et al, *Nature* (1977) 198:1056), the tryptophan (trp) promoter system (Goeddel et al, *Nuc Acids Res* (1980) 8:4057) and the lambda-derived  $P_L$  promoter and N gene ribosome binding site (Shimatake et al, *Nature* (1981) 292:128) and the hybrid tac promoter (De Boer et al, *Proc Nat Acad Sci USA* (1983) 292:128) derived from sequences of the trp and lac UV5 promoters. The foregoing systems are particularly compatible with *E. coli*; if desired, other prokaryotic hosts such as strains of *Bacillus* or *Pseudomonas*

20    30    may be used, with corresponding control sequences.

Eukaryotic hosts include without limitation yeast and mammalian cells in culture systems. Yeast expression hosts include *Saccharomyces*, *Klebsiella*, *Picia*, and the like. *Saccharomyces cerevisiae* and *Saccharomyces carlsbergensis* and *K. lactis* are the most commonly used yeast hosts, and are convenient fungal hosts.

Yeast-compatible vectors carry markers which permit selection of successful transformants by conferring prototrophy to auxotrophic mutants or resistance to heavy metals on wild-type strains. Yeast compatible vectors may employ the 2  $\mu$  origin of replication (Broach et al, *Meth Enzymol* (1983) 101:307), the combination of CEN3 and ARS1 or other means for assuring replication, such as sequences which will result in incorporation of an appropriate fragment into the host cell genome. Control sequences for yeast vectors are known in the art and include promoters for the synthesis of glycolytic enzymes (Hess et al, *J Adv Enzyme Reg* (1968) 7:149; Holland et al, *Biochem* (1978), 17:4900), including the promoter for 3-phosphoglycerate kinase (R. Hitzeman et al, *J Biol Chem* (1980) 255:2073). Terminators may also be included, such as those derived from the enolase gene (Holland, *J Biol Chem* (1981) 256:1385).

Mammalian cell lines available as hosts for expression are known in the art and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including HeLa cells, Chinese hamster ovary (CHO) cells, baby hamster kidney (BHK) cells, BSC 1 cells, CV1 cells, and a number of other cell lines. Suitable promoters for mammalian cells are also known in the art and include vital promoters such as that from Simian Virus 40 (SV40) (Fiers et al, *Nature* (1978) 273:113), Rous sarcoma virus (RSV), adenovirus (ADV), and bovine papilloma virus (BPV). Mammalian cells may also require terminator sequences and poly-A addition sequences. Enhancer sequences which increase expression may also be included, and sequences which promote amplification of the gene may also be desirable (for example methotrexate resistance genes). These sequences are known in the art.

Vectors suitable for replication in mammalian cells are known in the art, and may include vital replicons, or sequences which insure integration of the appropriate sequences encoding HCV epitopes into the host genome. For example, another vector used to express foreign DNA is Vaccinia virus. In this case the heterologous DNA is inserted into the Vaccinia genome and transcription can be directed by either endogenous vaccinia promoters or exogenous non-vaccinia promoters (e.g. T7 retroviral promoter) known to those skilled in the art, depending on the characteristics of the constructed vector. Techniques for the insertion of foreign DNA into the vaccinia virus genome are known in the art, and may utilize, for example, homologous recombination. The heterologous DNA is generally inserted into a gene which is non-

essential to the virus, for example, the thymidine kinase gene (tk), which also provides a selectable marker. Plasmid vectors that greatly facilitate the construction of recombinant viruses have been described (see, for example, Mackett et al, *J Virol* (1984) 49:857; Chakrabarti et al, *Mol Cell Biol* (1985) 5:3403; Moss, in *GENE*

5 TRANSFER VECTORS FOR MAMMALIAN CELLS (Miller and Calos, eds., Cold Spring Harbor Laboratory, N.Y., 1987), p. 10). Expression of the HCV polypeptide then occurs in cells or animals which are infected with the live recombinant vaccinia virus.

10 In order to detect whether or not the HCV polypeptide is expressed from the vaccinia vector, BSC 1 cells may be infected with the recombinant vector and grown on microscope slides under conditions which allow expression. The cells may then be acetone-fixed, and immunofluorescence assays performed using serum which is known to contain anti-HCV antibodies to a polypeptide(s) encoded in the region of the  
15 HCV genome from which the HCV segment in the recombinant expression vector was derived.

Other systems for expression of eukaryotic or viral genomes include insect cells and vectors suitable for use in these cells. These systems are known in the art,  
20 and include, for example, insect expression transfer vectors derived from the baculovirus *Autographa californica* nuclear polyhedrosis virus (AcNPV), which is a helper-independent, viral expression vector. Expression vectors derived from this system usually use the strong viral polyhedron gene promoter to drive expression of heterologous genes. Currently the most commonly used transfer vector for introducing  
25 foreign genes into AcNPV is pAc373 (see PCT WO89/046699 and U.S. Ser. No. 7/456,637). Many other vectors known to those of skill in the art have also been designed for improved expression. These include, for example, pVL985 (which alters the polyhedron start codon from ATG to ATT, and introduces a BamHI cloning site 32 bp downstream from the ATT; See Luckow and Summers, *Virol* (1989) 17:31). AcNPV  
30 transfer vectors for high level expression of non-fused foreign proteins are described in co-pending applications PCT WO89/046699 and U.S. Ser. No. 7/456,637. A unique BamHI site is located following position -8 with respect to the translation initiation codon ATG of the polyhedron gene. There are no cleavage sites for SmaI, PstI, BglII, XbaI or SstI. Good expression of non-fused foreign proteins usually requires foreign  
35 genes that ideally have a short leader sequence containing suitable translation

initiation signals preceding an ATG start signal. The plasmid also contains the polyhedron polyadenylation signal and the ampicillin-resistance (amp) gene and origin of replication for selection and propagation in *E. coli*.

5       Methods for the introduction of heterologous DNA into the desired site in the baculovirus virus are known in the art. (See Summer and Smith, Texas Agricultural Experiment Station Bulletin No. 1555; Smith et al, *Mol. Cell Biol.* (1983) 3:2156–2165; and Luckow and Summers, *Virol.* (1989) 17:31). For example, the heterologous DNA can be inserted into a gene such as the polyhedron gene by homologous  
10      recombination, or into a restriction enzyme site engineered into the desired baculovirus gene. The inserted sequences may be those which encode all or varying segments of the polyprotein, or other orfs which encode viral polypeptides. For example, the insert could encode the following numbers of amino acid segments from the polyprotein: amino acids 1–1078; amino acids 332–662; amino acids 406–662;  
15      amino acids 156–328, and amino acids 199–328.

The signals for post-translational modifications, such as signal peptide cleavage, proteolytic cleavage, and phosphorylation, appear to be recognized by insect cells. The signals required for secretion and nuclear accumulation also appear  
20      to be conserved between the invertebrate cells and vertebrate cells. Examples of the signal sequences from vertebrate cells which are effective in invertebrate cells are known in the art, for example, the human interleukin-2 signal (IL2<sub>S</sub>) which signals for secretion from the cell, is recognized and properly removed in insect cells.

25       Transformation may be by any known method for introducing polynucleotides into a host cell, including, for example packaging the polynucleotide in a virus and transducing a host cell with the virus, and by direct uptake of the polynucleotide. The transformation procedure used depends upon the host to be transformed. Bacterial transformation by direct uptake generally employs treatment with calcium or rubidium chloride (Cohen, *Proc. Nat. Acad. Sci. USA* (1972) 69:2110; T. Maniatis et at, “Molecular Cloning; A Laboratory Manual” (Cold Spring Harbor Press, Cold Spring Harbor, N.Y., 1982). Yeast transformation by direct uptake may be carried out using the method of Hinnen et al, *Proc. Nat. Acad. Sci. USA* (1978) 75:1929. Mammalian transformations by direct uptake may be conducted using the calcium phosphate  
30      precipitation method of Graham and Van der Eb, *Virol.* (1978) 52:546, or the various  
35      various

known modifications thereof. Other methods for introducing recombinant polynucleotides into cells, particularly into mammalian cells, include dextran-mediated transfection, calcium phosphate mediated transfection, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) 5 in liposomes, and direct microinjection of the polynucleotides into nuclei.

Vector construction employs techniques which are known in the art. Site-specific DNA cleavage is performed by treating with suitable restriction enzymes under conditions which generally are specified by the manufacturer of these 10 commercially available enzymes. In general, about 1 mg of plasmid or DNA sequence is cleaved by 1 unit of enzyme in about 20 mL buffer solution by incubation for 1–2 hr at 37° C. After incubation with the restriction enzyme, protein is removed by phenol/chloroform extraction and the DNA recovered by precipitation with ethanol. The cleaved fragments may be separated using polyacrylamide or agarose gel 15 electrophoresis techniques, according to the general procedures described in *Meth. Enzymol.* (1980) 65:499–560.

Sticky-ended cleavage fragments may be blunt ended using *E. coli* DNA polymerase I (Klenow fragment) with the appropriate deoxynucleotide triphosphates 20 (dNTPs) present in the mixture. Treatment with S1 nuclease may also be used, resulting in the hydrolysis of any single stranded DNA portions.

Ligations are carried out under standard buffer and temperature conditions using T4 DNA ligase and ATP; sticky end ligations require less ATP and less ligase 25 than blunt end ligations. When vector fragments are used as part of a ligation mixture, the vector fragment is often treated with bacterial alkaline phosphatase (BAP) or calf intestinal alkaline phosphatase to remove the 5'-phosphate, thus preventing re-ligation of the vector. Alternatively, restriction enzyme digestion of unwanted fragments can be used to prevent ligation. Ligation mixtures are transformed into 30 suitable cloning hosts, such as *E. coli*, and successful transformants selected using the markers incorporated (e.g., antibiotic resistance), and screened for the correct construction.

Synthetic oligonucleotides may be prepared using an automated 35 oligonucleotide synthesizer as described by Warner, *DNA* (1984) 3:401. If desired, the

synthetic strands may be labeled with  $^{32}\text{P}$  by treatment with polynucleotide kinase in the presence of  $^{32}\text{P}$ -ATP under standard reaction conditions.

DNA sequences, including those isolated from cDNA libraries, may be

5 modified by known techniques, for example by site directed mutagenesis (see e.g., Zoller, *Nuc. Acids Res.* (1982) 10:6487). Briefly, the DNA to be modified is packaged into phage as a single stranded sequence, and converted to a double stranded DNA with DNA polymerase, using as a primer a synthetic oligonucleotide complementary to the portion of the DNA to be modified, where the desired modification is included in

10 the primer sequence. The resulting double stranded DNA is transformed into a phage-supporting host bacterium. Cultures of the transformed bacteria which contain copies of each strand of the phage are plated in agar to obtain plaques. Theoretically, 50% of the new plaques contain phage having the mutated sequence, and the remaining 50% have the original sequence. Replicates of the plaques are hybridized to labeled

15 synthetic probe at temperatures and conditions which permit hybridization with the correct strand, but not with the unmodified sequence. The sequences which have been identified by hybridization are recovered and cloned.

DNA libraries may be probed using the procedure of Grunstein and Hogness

20 *Proc. Nat. Acad. Sci. USA* (1975) 73:3961. Briefly, in this procedure the DNA to be probed is immobilized on nitrocellulose filters, denatured, and pre-hybridized with a buffer containing 0–50% formamide, 0.75M NaCl, 75 mM Na citrate, 0.02% (wt/v) each of bovine serum albumin, polyvinylpyrrolidone, and Ficoll®, 50 mM  $\text{NaH}_2\text{PO}_4$  (pH 6.5), 0.1% SDS, and 100 m g/mL carrier denatured DNA. The percentage of

25 formamide in the buffer, as well as the time and temperature conditions of the pre-hybridization and subsequent hybridization steps depend on the stringency required. Oligomeric probes which require lower stringency conditions are generally used with low percentages of formamide, lower temperatures, and longer hybridization times. Probes containing more than 30 or 40 nucleotides, such as those derived from cDNA

30 or genomic sequences generally employ higher temperatures, e.g., about 40°–42° C., and a high percentage formamide, e.g., 50%. Following pre-hybridization, 5'- $^{32}\text{P}$ -labeled oligonucleotide probe is added to the buffer, and the filters are incubated in this mixture under hybridization conditions. After washing, the treated filters are subjected to autoradiography to show the location of the hybridized probe; DNA in

corresponding locations on the original agar plates is used as the source of the desired DNA.

For routine vector constructions, ligation mixtures are transformed into *E. coli* strain HB101 or other suitable hosts, and successful transformants selected by antibiotic resistance or other markers. Plasmids from the transformants are then prepared according to the method of Clewell et al, *Proc. Nat. Acad. Sci. USA* (1969) 62:1159, usually following chloramphenicol amplification (Clewell, *J. Bacteriol.* (1972) 110:667). The DNA is isolated and analyzed, usually by restriction enzyme analysis and/or sequencing. Sequencing may be performed by the dideoxy method of Sanger et al, *Proc. Nat. Acad. Sci. USA* (1977) 74:5463, as further described by Messing et al, *Nuc. Acids Res.* (1981) 9:309, or by the method of Maxam et al, *Meth. Enzymol.* (1980) 65:499. Problems with band compression, which are sometimes observed in GC-rich regions, were overcome by use of T-deazoguanosine according to Barr et al, *Biotechniques* (1986) 4:428.

Target plasmid sequences are replicated by a polymerizing means which utilizes a primer oligonucleotide to initiate the synthesis of the replicate chain. The primers are selected so that they are complementary to sequences of the plasmid. Oligomeric primers which are complementary to regions of the sense and antisense strands of the plasmids can be designed from the plasmid sequences already known in the literature.

The primers are selected so that their relative positions along a duplex sequence are such that an extension product synthesized from one primer, when it is separated from its template (complement), serves as a template for the extension of the other primer to yield a replicate chain of defined length.

The primer is preferably single stranded for maximum efficiency in amplification, but may alternatively be double stranded. If double stranded, the primer is first treated to separate its strands before being used to prepare extension products. Preferably, the primer is an oligodeoxyribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the agent for polymerization. The exact lengths of the primers will depend on many factors, including temperature and source of the primer and use of the method. For

example, depending on the complexity of the target sequence, the oligonucleotide primer typically contains about 15–45 nucleotides, although it may contain more or fewer nucleotides. Short primer molecules generally require cooler temperatures to form sufficiently stable hybrid complexes with the template.

5

The primers used herein are selected to be “substantially” complementary to the different strands of each specific sequence to be amplified. Therefore, the primers need not reflect the exact sequence of the template, but must be sufficiently complementary to selectively hybridize with their respective strands. For example, a 10 non-complementary nucleotide fragment may be attached to the 5'-end of the primer, with the remainder of the primer sequence being complementary to the strand. Alternatively, non-complementary bases or longer sequences can be interspersed into the primer, provided that the primer has sufficient complementarity with the sequence of one of the strands to be amplified to hybridize therewith, and to thereby form a 15 duplex structure which can be extended by the polymerizing means. The non-complementary nucleotide sequences of the primers may include restriction enzyme sites. Appending a restriction enzyme site to the end(s) of the target sequence would be particularly helpful for cloning of the target sequence.

20 It will be understood that “primer”, as used herein, may refer to more than one primer, particularly in the case where there is some ambiguity in the information regarding the terminal sequence(s) of the target region to be amplified. Hence, a “primer” includes a collection of primer oligonucleotides containing sequences representing the possible variations in the sequence or includes nucleotides which 25 allow a typical basepairing.

The oligonucleotide primers may be prepared by any suitable method. Methods for preparing oligonucleotides of specific sequence are known in the art, and include, for example, cloning and restriction of appropriate sequences, and direct 30 chemical synthesis. Chemical synthesis methods may include, for example, the phosphotriester method described by Narang et al. (1979), the phosphodiester method disclosed by Brown et al. (1979), the diethylphosphoramide method disclosed in Beaucage et al. (1981), and the solid support method in U.S. Pat. No. 4,458,066. The primers may be labeled, if desired, by incorporating means 35 detectable by spectroscopic, photochemical, biochemical, immunochemical, or

chemical means.

Template-dependent extension of the oligonucleotide primer(s) is catalyzed by a polymerizing agent in the presence of adequate amounts of the four  
5 deoxyribonucleotide triphosphates (dATP, dGTP, dCTP and dTTP) or analogs, in a reaction medium which is comprised of the appropriate salts, metal cations, and pH buffering system. Suitable polymerizing agents are enzymes known to catalyze primer- and template-dependent DNA synthesis. Known DNA polymerases include, for example, *E. coli* DNA polymerase I or its Klenow fragment, T<sub>4</sub> DNA polymerase,  
10 and Taq DNA polymerase. The reaction conditions for catalyzing DNA synthesis with these DNA polymerases are known in the art.

The products of the synthesis are duplex molecules consisting of the template strands and the primer extension strands, which include the target sequence. These  
15 products, in turn, serve as template for another round of replication. In the second round of replication, the primer extension strand of the first cycle is annealed with its complementary primer; synthesis yields a "short" product which is bounded on both the 5'- and the 3'-ends by primer sequences or their complements. Repeated cycles of denaturation, primer annealing, and extension result in the exponential  
20 accumulation of the target region defined by the primers. Sufficient cycles are run to achieve the desired amount of polynucleotide containing the target region of nucleic acid. The desired amount may vary, and is determined by the function which the product polynucleotide is to serve.

25 The PCR method can be performed in a number of temporal sequences. For example, it can be performed step-wise, where after each step new reagents are added, or in a fashion where all of the reagents are added simultaneously, or in a partial step-wise fashion, where fresh reagents are added after a given number of steps.  
30

In a preferred method, the PCR reaction is carried out as an automated process which utilizes a thermostable enzyme. In this process the reaction mixture is cycled through a denaturing region, a primer annealing region, and a reaction region. A machine may be employed which is specifically adapted for use with a thermostable  
35 enzyme, which utilizes temperature cycling without a liquid handling system, since the

enzyme need not be added at every cycle. This type of machine is commercially available from Perkin Elmer Cetus Corp.

After amplification by PCR, the target polynucleotides are detected by hybridization with a probe polynucleotide which forms a stable hybrid with that of the target sequence under stringent to moderately stringent hybridization and wash conditions. If it is expected that the probes will be completely complementary (i.e., about 99% or greater) to the target sequence, stringent conditions will be used. If some mismatching is expected, for example if variant strains are expected with the result that the probe will not be completely complementary, the stringency of hybridization may be lessened. However, conditions are chosen which rule out nonspecific/adventitious binding. Conditions which affect hybridization, and which select against nonspecific binding are known in the art, and are described in, for example, Maniatis et al. (1982). Generally, lower salt concentration and higher temperature increase the stringency of binding. For example, it is usually considered that stringent conditions are incubation in solutions which contain approximately 0.1×SSC, 0.1% SDS, at about 65° C. incubation/wash temperature, and moderately stringent conditions are incubation in solutions which contain approximately 1–2×SSC, 0.1% SDS and about 50°–65° C. incubation/wash temperature. Low stringency conditions are 2×SSC and about 30°–50°C.

Probes for plasmid target sequences may be derived from well known restriction sites. The plasmid probes may be of any suitable length which span the target region, but which exclude the primers, and which allow specific hybridization to the target region. If there is to be complete complementarity, i.e., if the strain contains a sequence identical to that of the probe, since the duplex will be relatively stable under even stringent conditions, the probes may be short, i.e., in the range of about 10–30 base pairs. If some degree of mismatch is expected with the probe, i.e., if it is suspected that the probe will hybridize to a variant region, the probe may be of greater length, since length seems to counterbalance some of the effect of the mismatch(es).

The probe nucleic acid having a sequence complementary to the target sequence may be synthesized using similar techniques described supra. for the

synthesis of primer sequences. If desired, the probe may be labeled. Appropriate labels are described supra.

In some cases, it may be desirable to determine the length of the PCR product  
5 detected by the probe. This may be particularly true if it is suspected that variant plasmid products may contain deletions within the target region, or if one wishes to confirm the length of the PCR product. In such cases it is preferable to subject the products to size analysis as well as hybridization with the probe. Methods for determining the size of nucleic acids are known in the art, and include, for example,  
10 gel electrophoresis, sedimentation in gradients, and gel exclusion chromatography.

The presence of the target sequence in a biological sample is detected by determining whether a hybrid has been formed between the polynucleotide probe and the nucleic acid subjected to the PCR amplification technique. Methods to detect  
15 hybrids formed between a probe and a nucleic acid sequence are known in the art. For example, for convenience, an unlabeled sample may be transferred to a solid matrix to which it binds, and the bound sample subjected to conditions which allow specific hybridization with a labeled probe; the solid matrix is then examined for the presence of the labeled probe. Alternatively, if the sample is labeled, the unlabeled  
20 probe is bound to the matrix, and after the exposure to the appropriate hybridization conditions, the matrix is examined for the presence of label. Other suitable hybridization assays are described supra. Analysis of the nucleotide sequence of the target region(s) may be by direct analysis of the PCR amplified products. A process for direct sequence analysis of PCR amplified products is described in Saiki et al.  
25 (1988).

Alternatively, the amplified target sequence(s) may be cloned prior to sequence analysis. A method for the direct cloning and sequence analysis of enzymatically amplified genomic segments has been described by Scharf (1986). In  
30 the method, the primers used in the PCR technique are modified near their 5'-ends to produce convenient restriction sites for cloning directly into, for example, an M13 sequencing vector. After amplification, the PCR products are cleaved with the appropriate restriction enzymes. The restriction fragments are ligated into the M13 vector, and transformed into, for example, a JM 103 host, plated out, and the resulting

plaques are screened by hybridization with a labeled oligonucleotide probe. Other methods for cloning and sequence analysis are known in the art.

Construction of the HCV/SEAP reporter gene plasmid

5

**General Method**

In the first embodiment, the Tropix® pCMV/SEAP expression vector is used as a starting point for construction of the HCV NS3 protease plasmid construct pHCAP1

10 (Seq. ID. NOS. 1-7). pHCAP1 is constructed from the pTM3 vector (Moss et al., *Nature*, 348:91-92 (1990)) in which the nucleotide sequence encoding the portion of the HCV-BK polyprotein domains NS2-NS3-NS4A-NS4B was cloned from the pBKCMV/NS2-NS3-NS4A-NS4B-SEAP (the pBK/HCAP) construct. pBK/HCAP is the eukaryotic expression plasmid in which all the original subcloning and ligation of all  
15 the HCV NS gene fragments and SEAP gene was created in. pCMV/SEAP is a mammalian expression vector designed for studies of promoter/enhancer elements with SEAP as a reporter (Berger et al., (1988)). The vector contains a polylinker for promoter/enhancer insertion, as well as an intron and polyadenylation signals from SV40. The vector can be propagated in *E.coli* due to the pUC19 derived origin of  
20 replication and ampicillin resistance gene. Modification of the commercially available plasmids is accomplished by use of PCR techniques including mutational PCR. Although this particular plasmid is described in the examples that follow, it is not the only plasmid or vector which may be used. The T7 RNA polymerase promoter is part of the pTM3 plasmid which was preferred in construction of the pHCAP vector.

25

In an alternate embodiment, the pTKgptF2s plasmid (Falkner and Moss, *J. Virol.* 62:1849-1854 (1988)) can be used instead of the pTM3 plasmid, which places the HCV/SEAP gene construct under transcriptional control of the native vaccinia virus promoter. The only requirement is that the promoter operate when placed in a  
30 plasmid having vaccinia virus regions flanking the subcloning region. This requirement allows the plasmid homologous recombination with the wild type vaccinia virus. Other vaccinia virus intermediate plasmids would be operable here as well.

**Example 1**

35 The Tropix® pCMV/SEAP expression vector is first modified so that both Sac1

restriction sites are inactivated. This is done by cleaving the plasmid with BamH1 which results in a 5' cleavage product that contains the plasmid 5' ATG site and about 250 bp ending at the Bam H1 site, and a 3' cleavage product having BamH1 sites at its 5' end and at its 3' end. The 5' cleavage fragment was then amplified from the 5 pCMV/SEAP plasmid using primers that were designed to delete the 5' ATG codon and to create a Sac 1 site on the 5' end. The downstream 3' primer spanned the Bam H1 site that is present within the SEAP coding sequence. Thus after PCR, the 10 amplified 5' fragment has a 5' Sac 1 site and a Bam H1 site. The 5' primer introduced an extra codon (a glutamic acid residue) in front of the first leucine residue of the 15 SEAP secretion signal. Furthermore, the first leucine codon was changed from a CTG to a CTC codon (a silent change). The codon change was made to create the second half of the Sac 1 site:

5'-GAGCTC-X-GGATCC-3' (Seq. ID NO:22)

15 Sac 1 site 5' end of SEAP Bam H1

The modified sequence is then cloned into pGEM3Zf(+) (Promega). The Bam H1-Bam H1 SEAP fragment was subcloned into pAlter-1 (Promega) which is a 20 plasmid that has an f1 origin of replication so it produces a single strand DNA for use in oligo mediated site directed mutagenesis. The Sac 1 sites within the SEAP fragment were mutated by oligo mediated site directed mutagenesis (GAGCTC to GAGCTG – a silent change) and the same change at the second Sac 1 site (GAGCTC to GAGCTG – an amino acid change from Serine to Cysteine) The 25 complete SEAP pGEM3Zf(+) plasmid is then made by subcloning the PCR modified 5' SEAP fragment into the Sac I- Bam H1 sites of pGEM3Zf(+). The resulting plasmid was then linearized with Bam H1 to allow the subcloning of the 3' SEAP Bam H1-Bam H1 from the pAlter-1 plasmid which was used for the oligo mediated site directed 30 mutagenesis to disrupt the two internal Sac I sites. A clone with the correct orientation of the Bam H1- Bam H1 fragment distal to the 5' SEAP fragment was selected after of purified plasmid DNA by restriction enzyme digest. This clone was used in the subsequent subcloning steps for the construction of the HCV/SEAP construct.

The coding sequences for the HCV proteins and NS3 cleavage sites that 35 comprise the final HCV/SEAP polyprotein were generated in two separate PCRs from

cDNA of the HCV-BK strain (Accession No. M58335). Takamizawa, A., et al., J. Virol. 65:1105-1113 1991. The first amplified fragment starts with the amino acid coding sequence of the HCV polyprotein corresponding to the C-terminal 81 amino acids of the putative E2 region, which are upstream of the beginning of the putative NS2

5 region or amino acid 729

(ARVCACLWMMLLIAQAEAALENLVLNSASVAGAHGILSFLVFFCAAWYIKGRLVPG  
ATYALYGVWPLLLLLALPPRAYAMDREMAA) (Seq. ID NO:23)

10 or nucleotide 2187

(GCACGTGTCTGTGCCTGCTTGTGGATGATGCTGCTGATAGCCCAGGCCGAGGC  
CGCCTTGGAGAACCTGGTGGCCTCAATGCGCGTCTGTGGCCGGCGCACATG  
GCATCCTCTCCTCCTTGTTCTCTGTGCCGCCTGGTACATCAAAGGCAGGCT  
15 GGTCCCTGGGGCGGCATATGCTCTTATGGCGTGTGGCCGCTGCTCCTGCTCTT  
GCTGGCATTACCACCGCGAGCTTACGCCATGGACCAGGAGATGGC) (Seq. ID  
NO:24)

and contains the DNA encoding the HCV polyprotein domains NS2-NS3-NS4A  
20 through the first 176 amino acids of the NS4B gene

(CASHLPYIEQ GMQLAEQFKQ KALGLLQTAT KQAEAAAPVV ESKWRALETF  
WAKHMWNFIS GIQYLAGLST LPGNPAIASL MAFTASITSPLTTQSTLLFN  
ILGGWVAAGL APPSAASAFV GAGIAGAAVG SIGLGKVLD  
25 ILAGYGAGVAGALVAFKVMS GEMPSTEDLV NLLPAIL) (Seq. ID NO:25)

or amino acid 1886 or nucleotide 5658

(TGCACCTCGCACCTCCCTTACATCGAGCAGGGAATGCAGCTGCCGAGCAATT  
30 CAAGCAGAAAGCGCTGGGTTACTGCAAACAGCCACCAACAAGCGGAGGCTG  
CTGCTCCGTGGTGGAGTCCAAGTGGCGAGCCCTTGAGACATTCTGGCGAAG  
CACATGTGGAATTTCATCAGCGGGATAACAGTACTTAGCAGGCTATCCACTCTGC  
CTGGGAACCCCGCAATAGCATCATTGATGGCATTACAGCCTCTATCACCAGCC  
CGCTCACCAACCAAAGTACCCCTCTGTTAACATCTGGGGGGTGGTGGCTG

CCCAACTCGCCCCCCCCAGCGCCGCTTCGGCTTCGTGGCGCCGGCATCGCC  
GGTGC GGCTTTGGCAGCATAGGCCTGGGAAGGTGCTTGACATTCTGGC  
GGGTTATGGAGCAGGAGTGGCCGGCGCGCTCGTGGCCTTAAGGTCATGAGCG  
GCGAGATGCCCTCCACCGAGGACCTGGTCAATCTACTTCCTGCCATC) (Seq. ID

5 NO:26)

The primers used to amplify the fragment were designed to contain an Eco RI site and an ATG codon in the 5' primer (Seq. ID NO:27) and an Xho I site in the 3' primer (Seq. ID NO:28). The amplified fragment was accordingly subcloned as an Eco RI -  
10 Xho I fragment into pET24a(+) plasmid (Novagen). The second fragment amplified from the HCV strain BK cDNA encompasses the putative NS5A/5B cleavage site (EEASEDVVCCSMSYWTGAL)(Seq. ID NO:29). The 5' primer that was used to amplify the cleavage site was designed to have an Xho I site (Seq. ID NO:30) whereas the 3' primer was designed to have a Sac I site (Seq. ID NO:31). The  
15 resulting PCR product was subcloned as an Xho I - Sac I fragment into pET24a(+), which had been digested with Xho I- Hind III, as part of a three way ligation (Seq. ID NO:32). The third fragment in the three way ligation was the Sac I - Hind III fragment from the SEAP pGEM3Zf(+) plasmid. The Sac I - Hind III fragment encompassed the modified SEAP gene and also 30 base pairs of the pGEM3Zf(+) polylinker which  
20 included the multiple cloning sites (MCS) between the Bam H1 and HindIII sites. The final HCV/SEAP construct was assembled using pBKCMV as the vector. pBKCMV was digested with Eco RI and Hind III and then used in a three way ligation with the NS5A/5B - SEAP Xho I -Hind III fragment and the Eco RI-Xho I NS2-NS4B fragment.

25 The control plasmids for the assay (pHCAP3, pHCAP4) were constructed in a similar manner to the HCV/SEAP construct. The control plasmids have either an inactive form of NS3 protease or inactive forms of both NS2 protease and NS3 protease. Inactivation of NS2 and NS3 proteases was accomplished by oligo mediated site directed mutagenesis performed on the PCR amplified NS2 - NS4B  
30 fragment that had been subcloned into pALTER-1 as an Eco R1 - Xho 1 fragment together with the NS5A/5B Xho 1 - Sac 1 fragment. In order to inactivate the NS3 protease, the catalytic serine residue was substituted with an alanine by replacing thymidine (TCG) with guanine (GCG)(base 2754). The NS2 protease was inactivated by substitution of the catalytic cysteine residue with an alanine residue (TGT ->  
35 GCT)(bases 2238-2239). The resulting inactivated NS3 protease and inactivated

NS2-NS3 proteases variants of the NS2-NS4B fragment were each subcloned into pBKCMV as separate Eco R1 - Xho 1 fragments together with the NS5A/5B - SEAP Xho 1 - Hind III fragment.

5        The pHCAP1 ( $\text{NS2}^{\text{WT}}\text{NS3}^{\text{WT}}$ ) (Seq. ID NOS:1-7), pHCAP3 ( $\text{NS2}^{\text{WT}}\text{NS3}^{\text{MUT}}$ ) (Seq. ID NOS:8-14), and pHCAP4 ( $\text{NS2}^{\text{MUT}}\text{NS3}^{\text{MUT}}$ ) (Seq. ID NOS:15-21) plasmids were constructed using pTM3 as the vector and the appropriate HCV/SEAP fragment from the corresponding pBKHCV/SEAP constructs. The pBKHCV/SEAP constructs were first digested with Eco R1 and the Eco R1 site was filled in using  
10 Klenow fragment in a standard fill in reaction. The pBKHCV/SEAP constructs were then digested with Xba I and the gel purified HCV/SEAP fragment was subcloned into pTM3 that had been digested with Sma 1 and Spe 1. Subcloning the HCV/SEAP fragment into the Sma I site will result in an additional 6 amino acids (MGIPQF) (Seq. ID NO:33) at the N-terminus (codons 1426-1444) if the preferred translational start  
15 codon, which is part of the Nco 1 site in pTM3, is used.

The pHCAP1 ( $\text{NS2}^{\text{WT}}\text{NS3}^{\text{WT}}$ ), pHCAP3 ( $\text{NS2}^{\text{WT}}\text{NS3}^{\text{MUT}}$ ), and pHCAP4 ( $\text{NS2}^{\text{MUT}}\text{NS3}^{\text{MUT}}$ ) plasmids have been used to generate recombinant vaccinia viruses as described in the next section.

20        Construction of the HCV/SEAP reporter gene viral vectors

Applicants have generated recombinant vaccinia virus using pHCAP1 and the control plasmids, pHCAP3 and pHCAP4. Recombinant vaccinia viruses were  
25 generated using standard procedures in which BSC-1 cells were infected with wild type vaccinia virus (strain WR from ATCC) and then transfected with either pHCAP1, pHCAP3, or pHCAP4. Selection of recombinant virus was performed by growth of infected transfected cells in the presence of mycophenolic acid. The recombinant vaccinia viruses are termed vHCAP1, vHCAP3, and vHCAP4 and correspond directly  
30 with the pHCAP1, pHCAP3, and pHCAP4 plasmids. Large scale stocks of the vHCAP1, vHCAP3, and vHCAP4 were grown and titered in CV1 cells.

Transfection of Cell Lines Containing the HCV/SEAP reporter

In the first embodiment HeLa cells are transfected with the Hep C/SEAP reporter gene plasmid, pHCAP1, and co-infection with a vTF7.3, a recombinant vaccinia virus (Fuerst et al., *Proc. Nat. Acad. Sci. USA*, 86:8122-8126 (1986)). vTF7.3 expresses T7 RNA polymerase which is required for transcription of the reporter gene since it is under the control of T7 promoter in the pTM3 plasmid. The pTM3 plasmid is a vaccinia intermediate plasmid which can function as an expression vector in cells when T7 RNA polymerase is provided in *trans* (Figure 2).

As described previously, the Hep C/SEAP reporter gene encodes for a polyprotein with the following gene order: HCV (strain BK) NS2-NS3-NS4A-NS4B' - NS5A/5B cleavage site - SEAP. Thus the HCV sequences for the amino acid coding sequence of the HCV polyprotein corresponding to the C-terminal 81 amino acids of the putative E2 region, which are upstream of the start of the putative NS2 region (as defined by Grakoui et al. ) or amino acid 729 and continues through the first 176 amino acids of the NS4B gene or amino acid 1886 (Seq. ID NOS:23-26), and is proximal to the SEAP protein (see Figure 1). The NS5A/5B cleavage site has been engineered between the end of NS4B' and the second codon of SEAP.

The working theory behind the unique design of the reporter gene construct is that the SEAP polyprotein is tethered, as part of the NS2-NS3-NS4A-NS4B' - NS5A/5B cleavage site – SEAP polyprotein, inside the cell. It has been shown that NS2 is a hydrophobic protein and is associated with the outside of the endoplasmic reticulum (ER). Grakoui, et al. (1993). Thus, in the present invention, SEAP is tethered to the ER via the action of NS2. Release of SEAP from the polyprotein tether will occur upon NS3-mediated cleavage at the NS5A/5B cleavage site. SEAP is then secreted from the cell and can be monitored by assaying media for alkaline phosphatase activity (Figure 1B). It is assumed that it is NS3-mediated cleavage at the NS5A/5B site which is the necessary cleavage to release SEAP from the upstream polyprotein sequences. However NS3-mediated cleavage at other sites within the polyprotein may be responsible for SEAP release and hence its subsequent secretion. Both NS3 and NS3/NS4A, where NS4A is a cofactor for NS3, can mediate cleavage at the NS3/4A and NS4A/4B cleavage sites which are present in polyprotein in addition to the engineered NS5A/5B cleavage site. Thus there may be more than

one NS3-mediated cleavage event occurring over the length of the polyprotein before SEAP is available to the cell secretion apparatus and secreted from the cell. Further, in an alternative embodiments the tether may be changed depending upon the chosen cleavage site. In addition, NS2 is an autocatalytic protease; it mediates the 5 cleavage event between its carboxy-terminal end and the NS3 N-terminus. In the Hep C/SEAP polyprotein, NS2-mediated cleavage at the NS2/NS3 site would release the NS3-NS4A-NS4B'-SEAP polyprotein from the ER.

The above described system can be used to evaluate potent NS3 inhibitors by 10 monitoring the effect of increasing drug concentration on SEAP activity. NS3 inhibition would be detected as a decrease in SEAP activity. Recognizing that a decrease in SEAP activity could also be due to cell cytotoxicity of a given compound or a non-specific effect on vaccinia virus which would adversely effect SEAP transcription, appropriate controls are used as discussed below.

15

In an alternate embodiment, a "cis-only" cleavage assay is contemplated. In this assay the NS2<sup>MUT</sup>NS3<sup>WT</sup> variant of the HCV/SEAP (HCAP2) is used so the polyprotein remains tethered to the outside of the endoplasmic reticulum because the 20 NS2 protease cannot catalyze the cleavage between the C-terminus and the NS3 N-terminus. Thus the only way for SEAP to be released from the tether is if the NS3 protease clips in cis at the NS5A/5B cleavage site. There should not be any trans NS3 mediated cleavage events occurring since NS2 is not available to release the NS3 N-terminus from its tether. The control plasmid or virus for this assay is the 25 NS2<sup>MUT</sup>NS3<sup>MUT</sup> variant HCAP4.

25

#### DI/DR Assay

A preferred embodiment involves the co-infection of BHK (ATCC No. CCL-10) or CV1 cells (a COS1 derived line ATCC No. CCL-70) cells with both vHCAP1 and 30 vTF7.3 (ATCC No. VR-2153), with CV1 being more preferred. The latter virus is necessary since the Hep C/SEAP gene remains under control of the T7 RNA polymerase promoter in the vHCAP recombinant viruses. Currently both 35 embodiments which are termed the Hep C/SEAP transfection/infection assay, and the dual recombinant vaccinia virus assay (DI/DR assay) respectively, are useful for HCV protease candidate compound evaluation (Figure 3).

Example 1*Protocol for vTF7.3 infection / HCV/SEAP Plasmid Transfection Experiment*

## 5 Day 1

Flat-bottom 96 well plates were seeded with BHK cells at a density of  $1 \times 10^4$  cells/well (equivalent to about 85% confluence) after 24 hours. In general, one 96 well plate was used for investigation of each compound of interest (protease inhibitor), plus an additional plate at the same cell density is used where two rows are 10 designated for each compound of interest at increasing concentrations for investigating the cytotoxicity of the compounds themselves in cells alone. Cytotoxicity was determined by XTT assay (Sigma 4626).

## Day 2

15 The established monolayer was transfected with either pHCap1, pHCap3, pHCap4, or pTM3 plasmids at a concentration of 0.4  $\mu\text{g}/\text{well}$  as part of a DNA Lipofectamine (Gibco BRL) transfection mixture. Infections of the established monolayer with vTF7.3 preceded the transfection step. A working stock of vTF7.3 was diluted to a multiplicity of infection (MOI) of 10 with Optimem. The media was 20 aspirated from the wells (2B-10G) 2 rows at a time. A 50 L aliquot of vTF7.3 inoculum was added per well and gently shaken every 10 minutes. 30 minutes after inoculum addition, the transfection mixes were made by adding 1 mL of Optimem in 3 mL polystyrene tubes. To the media, 48  $\mu\text{g}$  of plasmid DNA was then added to the tubes and mixed, followed by 144  $\mu\text{L}$  of Lipofectamine<sup>TM</sup>, and then the mixture was 25 incubated (R.T.) for 30 minutes. After incubation, 11 mL of Optimem were added to each of the tubes and gently mixed. The vTF7.3 inoculum was aspirated from the wells and 0.1 mL of transfection mix was added to each well and incubated at 34 °C for 4 hours. Compounds/drugs of interest for testing protease inhibition were prepared as stock solutions of 40 mM in 100% DMSO. For assay use, the 30 compounds were diluted to 640  $\mu\text{M}$  (2X) in Optimem with 4% FBS. The compound dilutions were set up in an unused 96 well plate by adding 100  $\mu\text{L}$  Optimem with 4% FBS to wells 4-10 and 150  $\mu\text{L}$  of compound dilutions to all wells in column 3. A serial dilution of the compounds was then performed by transferring 46  $\mu\text{L}$  from well to well across the plate. The transfection mixture was then aspirated from the cells. Then 75

$\mu$ L of Optimem with 4% FBS was added to the transfected monolayers. Add 75  $\mu$ L of the 2X compound dilutions to the transfected monolayers and incubated at 34 °C for 48 hours. The cells were checked microscopically at 24 hours and media is collected at 48 hours for measurement of SEAP activity.

5

*SEAP Activity Measurement*

After 48 hours, SEAP activity was measured by first transferring 100  $\mu$ l of media from each well of the 96 well assay plate to a new sterile 96 well plate. Plate(s) 10 were sealed and heated in a heating block at 65 C for 30 minutes. After 30 minutes, plate(s) were removed and cooled to room temperature. For each heat treated plate, we transferred 50  $\mu$ l of heat treated media to a Dynex (Dynex 7416) 96 well plate. To each well was added 50  $\mu$ l of Tropix assay buffer and incubated at room temperature for 5 minutes, followed by an addition to each well of 50  $\mu$ l of Tropix reaction 15 buffer/CSPD substrate (Tropix), each was mixed, and incubated for an additional 90 minutes at room temperature. Chemiluminescence was read in the Victor multilabel counter from Wallac, Inc. (model number 1420) as one second counts and data is reported as luminescent units/second.

20 For Examples 1 and 2:

*XTT Cytotoxicity Assay*

XTT (Sigma 4626) was dissolved in phosphate buffered saline (PBS) to a final 25 concentration of 1 mg/mL. 5 mL was prepared per plate. To this solution was added 5 mM PMS (n-methyldibenzopyrazine methyl sulfate salt) (Sigma P9625) to a final concentration of 20  $\mu$ M. 50  $\mu$ L of the XTT solution was added per well to the plate set up for cytotoxicity. The plates were incubated at 37 C in a 5% CO<sub>2</sub> incubator for about 3.5 hours and then the color change was quantitated by reading absorbance in a Vmax plate 30 reader (Molecular Devices) at 450nm/650 nm. Values were corrected by subtracting media-only background and presented as %viable with the untreated cell control representing 100%.

Example 2

35

Representative experiment and resulting data using Protocol of Example 1.

Compounds X, Y, and Z were evaluated in the Vaccinia Virus Infection/

5 Plasmid Transfection assay as outlined in Example 1. BHK cells were seeded into 96 well plates at a density of  $1 \times 10^4$  cells/well and grown overnight to approximately 85% confluence. The SEAP activity was monitored 48 hours post drug addition in cells transfected with either pHCAP1, pHCAP4, pTM3, or no DNA. Concurrently, Compounds X, Y, and Z were evaluated for cell cytotoxicity in a separate dose

10 response assay using XTT to measure cell viability.

For each compound, cells were infected with vTF7.3 followed by the plasmid transfection step. The arrangement of the cells transfected with one of the three plasmids is illustrated in Figure 9.

15

Results for Compounds X, Y, and Z are shown in Figures 4 A and 4B and Table 1 below. In the three graphs, the amount of SEAP activity detected in cells transfected with the pHCAP1 plasmid ranges from 2 to 7-fold above the amount of

20 SEAP detected in cells transfected with the control plasmids, pHCAP4 and pTM3, or cells only. The EC<sub>50</sub> ( $\mu$ M) value represents the concentration of drug at which a 50% reduction in SEAP activity is observed relative to the amount of SEAP activity detected in the absence of drug. The CC<sub>50</sub> ( $\mu$ M) value represents the concentration of drug at which a 50% reduction in cell viability is observed relative to cells in the

25 absence of drug. The ratio of EC<sub>50</sub>/ CC<sub>50</sub> yields the therapeutic index (TI) which, by convention, should be greater or equal to 10 in order for a compound to be considered as demonstrating antiviral activity.

Table 1

30

Compound	EC <sub>50</sub> ( $\mu$ M)	CC <sub>50</sub> ( $\mu$ M)	Solubility ( $\mu$ M)	TI
X	45	178	= 100	4
Y	>320	112	= 100	-
Z	>320	112	= 100	-

Within the compound dose range that was examined, only an EC<sub>50</sub> value for Compound X was obtained. However, since the TI value for Compound X was below 10, it was concluded that Compound X does not represent a candidate inhibitor of NS3 protease activity. Compounds Y and Z did not demonstrate any efficacy in this system and, therefore, are not considered potential candidates (Figs. 4A and 4B).

For Examples 3 and 4:

10    *XTT Cytotoxicity Assay*

XTT (Sigma 4626) was dissolved in phosphate buffered saline (PBS) to a final concentration of 1 mg/mL. 5 mL were prepared per plate. To this solution was added 5 mM PMS (n-methyldibenzopyrazine methyl sulfate salt) (Sigma P9625) to a final concentration of 20 µM. This XTT substrate solution was diluted with an equal volume of MEM media containing 4% FBS(V/V). A 100µL/well of this final solution was added to the original plate which still contains the cell monolayer and about 50 µL incubation media. The plates were Incubated at 37 C in a 5% CO<sub>2</sub> incubator for about 3.5 hours and then the color change was quantitated by reading absorbance in a Vmax plate reader (Molecular Devices) at 450nm/650 nm. Values were corrected by subtracting media-only background and presented as %viable with the untreated cell control representing 100%.

Example 3

25    *Protocol for Dual Infection/Dose Response (DI/DR) Assay*

Day 1

Flat-bottom 96-well plates were seeded with CV1 cells at a density of 1 x 10<sup>5</sup> cells per well in MEM media containing 10% FBS with no Phenol Red. The plate was set up as shown in Figure 5. Media only was placed in all the wells on the edge of the plate and only one compound is evaluated per plate (Fig. 5).

Day 2

Cells were infected with recombinant vaccinia viruses as follows. There should be about  $1.5 \times 10^5$  cells per well after incubation for 24 hours. For every plate needed (a plate for each drug in the experiment) 4 mL of vTF7.3 in MEM with 4% FBS (-) phenol red at a concentration of  $2 \times 10^6$  pfu/mL was prepared, and divided into 2  
5 mL aliquots. Either vHCAP1 or vHCAP3 was added to the vTF7.3 aliquots for a final concentration of vHCAP of  $1 \times 10^7$  pfu/mL. At 75  $\mu$ L per well, this concentration of virus stock delivers vTF7.3 at an MOI of 1 and vHCAP1 or vHCAP3 at an MOI of 5. The arrangement of the experimental plate is shown in Figure 5.

10 Drug stock solutions for use in the assay, were made at a concentration of 40 mM in DMSO as in the previous protocol. The 40 mM drug stock solution was diluted to 640  $\mu$ M in MEM with 4% FBS (-) phenol red to yield a 2X drug working stock solution. Using an empty 96 well plate, the drug dilution series was set up as follows:

15 100  $\mu$ L of MEM with 4% FBS (-) phenol red was added to all wells in columns 4-10. 150  $\mu$ L of 2X drug working stock solution was added to all wells in column 3. 46  $\mu$ L of media was transferred from column 3 to wells of column 4 and mixed. Transferring of 46  $\mu$ L from column 4 to column 5 and out to row 10 was repeated. The remaining 46  $\mu$ L was discarded. The arrangement of the experimental multiwell  
20 plate is shown in Figure 6.

Media was aspirated from the CV1 monolayers. After aspiration, 75  $\mu$ L per well of appropriate virus inoculum or MEM with 4% FBS (-) phenol red was added to the CV1 monolayers, then 75  $\mu$ L was transferred from each well in the drug dilution  
25 series plate to the corresponding wells on the cell monolayer plate. The assay plate was incubated at 37 C in a 5% CO<sub>2</sub> incubator for 48 hours.

At Day 3, the cells was microscopically checked for phenotypic changes around the 24 hour time point. At Day 4, 100  $\mu$ L of media was collected from each  
30 well of which 50  $\mu$ L was used in the measurement of SEAP activity. The 100  $\mu$ L aliquots were transferred to an unused 96 well plate and after the plate was sealed, it was heated to 65 C for 30 minutes. 50  $\mu$ L of each heat treated sample was then transferred to its corresponding well in a new 96 well opaque plate (Dynex 7416). Using the Tropix® SEAP PhosphaLight™ kit, 50 mL of Tropix assay buffer was added

to each well and the plate was incubated at room temperature for 5 minutes. Next, 50  $\mu$ L of Tropix reaction buffer/CPSD substrate was added and mixed. The plate was incubated for 90 minutes at room temperature. The chemiluminescence was then read using a Victor multi-label counter. The XTT assay for measuring cytotoxicity was 5 also performed on Day 4 as described.

Example 4

*Representative Experiment and Resulting Data Using Protocol of Example 3*

10

Compounds A -I were evaluated in the DI/ DR assay using the standard protocol given in Example 3. The data shown in Figure 7 and Figure 8 represent assay results obtained at a 48 hour time point post drug addition.

15

The EC<sub>50</sub> ( $\mu$ M) value represents the concentration of drug at which a 50% reduction in SEAP activity is observed relative to the amount of SEAP activity detected in the absence of drug. However, this latter value, the amount of SEAP activity that is observed in the absence of drug, is first corrected for assay background prior to the calculation of an EC<sub>50</sub> value. The correction is made since in the inactive

20

NS3 protease construct, vHCAP3, a background level of SEAP activity is detected (see SEAP Activity graph). This background SEAP activity represents non-NS3 protease mediated SEAP activity and therefore should not be affected by the addition of an NS3 protease inhibitor. It is assumed that a fraction of the SEAP activity that is observed in the active NS3 protease construct, vHCAP1, represents non-NS3

25

protease mediated SEAP activity. Therefore the amount of SEAP activity detected vHCAP1 is corrected for the fraction that corresponds to non-NS3 protease mediated SEAP activity. The correction is as follows: luminescent units of SEAP activity of vHCAP1 - luminescent units of SEAP activity of vHCAP3 = Value N (level of NS3 protease dependent SEAP activity). Accordingly, (vHCAP1/SEAP)-N/2 = EC<sub>50</sub> value.

30

The CC<sub>50</sub> ( $\mu$ M) value represents the concentration of drug at which a 50% reduction in cell viability is observed relative to cells in the absence of drug. The ratio of EC<sub>50</sub>/ CC<sub>50</sub> yields the therapeutic index (TI) which, by convention, should be

greater or equal to 10 in order for a compound to be considered as demonstrating antiviral activity.

In Figure 7, increasing concentrations of Compound A were observed to have  
5 no affect on SEAP activity. In the cell cytotoxicity component of the assay, it was  
observed that increasing concentrations of Compound A did not result in a reduction  
of cell viability of cells alone or cells infected with either vHCAP1/vTF7.3 or  
vHCAP3/vTF7.3. The results obtained with Compounds B - I (Figure 8) demonstrate  
a range of observed cytotoxicities from 15  $\mu$ M to >320  $\mu$ M which is the upper limit of  
10 drug concentrations tested in the DI/ DR assay although it is theoretically possible to  
test drug concentrations above 320  $\mu$ M. The EC<sub>50</sub> values that were observed for  
Compounds B - I ranged from 18  $\mu$ M to > 320  $\mu$ M, however, the TI values were under  
10. Thus Compounds A -I do not represent potential inhibitors of NS3 protease  
activity.

We Claim:

1. A reporter gene system useful in the assessment of compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:
  - a) a recombinant viral vector comprising a DNA molecule encoding an RNA polymerase promoter compatible with said viral vector and which is expressed upon infection of a target mammalian cell;
  - b) a recombinant plasmid comprising a DNA molecule encoding the HCV/SEAP reporter gene polyprotein which is expressed when transfected into a target mammalian cell;
  - c) said target mammalian cell line being infected first with said recombinant viral vector then transfected with said recombinant plasmid such that the DNA molecule encoding the HCV/SEAP reporter gene is under transcriptional control of said promoter; and
  - d) the target mammalian cell expressing said HCV/SEAP reporter gene polyprotein such that SEAP is secreted from said target mammalian cell.
2. A reporter gene system useful in the assessment of compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:
  - a) a first recombinant viral vector comprising a DNA molecule encoding an RNA polymerase promoter compatible with said viral vector and which is expressed upon infection of a target mammalian cell;
  - b) a second recombinant viral vector comprising a DNA molecule encoding the HCV/SEAP reporter gene polyprotein which is expressed upon infection of a target mammalian cell;
  - c) said target mammalian cell line being infected first with said first

recombinant viral vector then co-infected with said second recombinant plasmid such that the DNA molecule encoding the HCV/SEAP reporter gene is under control of said promoter; and

- d) the target mammalian cell expresses said HCV/SEAP reporter gene polyprotein such that SEAP is secreted from said target mammalian cell.

3. The reporter gene system of claim 1 wherein said recombinant plasmid is the pTM3 plasmid containing said HepC/SEAP construct.
4. The recombinant plasmid of claim 3 wherein said recombinant plasmid comprises the pHCP1 plasmid having a DNA molecule encoding the NS2 and NS3 protease polyproteins in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 1.
5. The recombinant plasmid of claim 3 wherein said recombinant plasmid further comprises the pHCP3 plasmid containing the active NS2 protease and a mutant NS3 protease in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 8.
6. The recombinant plasmid of claim 3 wherein said recombinant plasmid further comprises the pHCP4 plasmid containing the mutant inactive NS2 and mutant inactive NS3 protease in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 15.
7. The reporter gene system of claim 2 wherein said second recombinant viral vector further comprises the vHCP1 vector having a DNA molecule encoding the NS2 and NS3 protease polyproteins in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 1.
8. The reporter gene system of claim 2 wherein said second recombinant viral vector further comprises the vHCP3 vector containing the active NS2 protease and a mutant NS3 protease in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 9.

9. The reporter gene system of claim 2 wherein said second recombinant viral vector further comprises the vHCAP4 vector containing the active NS2 protease and a mutant NS3 protease in a fusion protein fused with the SEAP gene according to the sequence in Seq. ID NO: 16.
10. The reporter gene system of claim 1 wherein said recombinant viral vector comprises a virus containing the DNA sequence encoding T7 RNA polymerase promoter.
11. The recombinant viral vector of claim 7 wherein said vector is the vTF7.3 vector.
12. The reporter gene system of claim 2 wherein said first recombinant viral vector comprises a virus containing the DNA sequence encoding the T7 RNA polymerase promoter.
13. The recombinant viral vector of claim 9 wherein said vector is the vTF7.3 vector.
14. The reporter gene system of claim 1 wherein said first recombinant viral vector comprises a virus containing the DNA sequence encoding a vaccinia virus compatible promoter.
15. The first recombinant viral vector of claim 11 wherein said vector is a vaccinia virus derived vector.
16. The reporter gene system of claim 2 wherein said first recombinant viral vector comprises a virus containing the DNA sequence encoding a vaccinia virus compatible promoter.
17. The first recombinant viral vector of claim 13 wherein said vector is a vaccinia virus derived vector.

18. A first recombinant viral vector according to claim 2 wherein the vector is pTM3 plasmid, a Listeria vector, an orthopox virus, avipox virus, canarypox virus, suipox virus, vaccinia virus, baculovirus, human adenovirus, SV40, Herpes Virus or bovine papilloma virus.
19. A second recombinant viral vector according to claim 2 wherein the vector is pTM3 plasmid, a Listeria vector, an orthopox virus, avipox virus, canarypox virus, suipox virus, vaccinia virus, baculovirus, human adenovirus, SV40, Herpes Virus or bovine papilloma virus.
20. The reporter gene system of claim 1 wherein said recombinant viral vector comprises a virus containing a the DNA sequence encoding a promoter selected from the group of mammalian viral vectors consisting of:  
  
Simian Virus 40 (SV40), Rous Sarcoma Virus (RSV), Adenovirus (ADV) and Bovine Papilloma Virus (BPV).
21. The reporter gene system of claim 2 wherein said recombinant viral vector comprises a virus containing a the DNA sequence encoding a promoter selected from the group of mammalian viral vectors consisting of:  
  
Simian Virus 40 (SV40), Rous Sarcoma Virus (RSV), Adenovirus (ADV) and Bovine Papilloma Virus (BPV).
22. The reporter gene system of claim 1 wherein said target cell line is selected from the group consisting of:  
  
HeLa cells, Chinese Hamster Ovary cells, CV1 African Green Monkey cells, BSC 1 cells and Baby Hamster Kidney cells.
23. The reporter gene system of claim 2 wherein said target cell line is selected from the group consisting of:

HeLa cells, Chinese Hamster Ovary cells, CV1 African Green Monkey cells, BSC 1 cells and Baby Hamster Kidney cells.

24. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the HepC/SEAP reporter gene construct according to claim 1.
25. The isolated DNA sequence of claim 24 comprising a DNA sequence or variants thereof in SEQ. ID NO. 1.
26. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as pHCAP1.
27. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as pHCAP3.
28. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as pHCAP4.
29. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as vHCAP1.
30. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as vHCAP3.
31. An isolated DNA sequence comprising a DNA sequence or variants thereof encoding the sequence defined as vHCAP4.
32. A method of assessing compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:
  - a) a control target mammalian cell;
  - b) a first target mammalian cell expressing the pHCAP1 polyprotein;
  - c) a second target mammalian cell expressing the pHCAP4 polyprotein;

- d) a third target mammalian cell expressing the viral promoter only;
- e) incubating said control, first, second, and third target mammalian cells for about 24 hours in a suitable growth medium in the presence and/or absence of pharmacologically effective concentrations of candidate compounds;
- f) measuring the amount of SEAP activity; and
- g) determining whether said candidate compounds augmented or inhibited hepatitis C NS3 protease by comparing the SEAP activity of said control, first, second, and third target mammalian cells.

33. A method of assessing compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:

- a) a control target mammalian cell;
- b) a first target mammalian cell expressing the vHCAP1 polyprotein;
- c) a second target mammalian cell expressing the vHCAP4 polyprotein;
- d) a third target mammalian cell expressing the viral promoter only;
- e) incubating said control, first, second, and third target mammalian cells for about 24 hours in a suitable growth medium in the presence and/or absence of pharmacologically effective concentrations of candidate compounds;
- f) measuring the amount of SEAP activity; and
- g) determining whether said candidate compounds augmented or inhibited hepatitis C NS3 protease by comparing the SEAP activity of said control, first, second, and third target mammalian cells.

34. A method of assessing compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease cis-only cleavage comprising:

- a) a control target mammalian cell;
- b) a first target mammalian cell expressing the pHCap3 polyprotein;
- c) a second target mammalian cell expressing the pHCap4 polyprotein;
- d) a third target mammalian cell expressing the viral promoter only;
- e) incubating said control, first, second, and third target mammalian cells for about 24 hours in a suitable growth medium in the presence and/or absence of pharmacologically effective concentrations of candidate compounds;
- f) measuring the amount of SEAP activity; and
- g) determining whether said candidate compounds augmented or inhibited hepatitis C NS3 protease by comparing the SEAP activity of said control, first, second, and third target mammalian cells.

35. A process for constructing a reporter gene system useful in the assessment of compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:

- a) providing a recombinant viral vector comprising a DNA molecule encoding an RNA polymerase promoter compatible with said viral vector and which is expressed upon infection of a target mammalian cell;
- b) providing a recombinant plasmid comprising a DNA molecule encoding the HCV/SEAP reporter gene polyprotein which is expressed when transfected into a target mammalian cell further comprising the steps of

cloning into a suitable vector the NS2-NS3-NS4A-NS4B' -NS5A/5B cleavage site – SEAP polyprotein;

- c) said target mammalian cell line being infected first with said recombinant viral vector then transfected with said recombinant plasmid such that the DNA molecule encoding the HCV/SEAP reporter gene is under transcriptional control of said promoter; and
- d) the target mammalian cell expressing said HCV/SEAP reporter gene polyprotein such that SEAP is secreted from said target mammalian cell.

36. A process for constructing a reporter gene system useful in the assessment of compounds which augment or inhibit the activity of Hepatitis C virus NS3 protease comprising:

- a) providing a first recombinant viral vector comprising a DNA molecule encoding an RNA polymerase promoter compatible with said viral vector and which is expressed upon infection of a target mammalian cell;
- b) providing a second recombinant viral vector comprising a DNA molecule encoding the HCV/SEAP reporter gene polyprotein which is expressed when transfected into a target mammalian cell further comprising the steps of cloning into a suitable vector the NS2-NS3-NS4A-NS4B' -NS5A/5B cleavage site – SEAP polyprotein;
- c) said target mammalian cell line being infected first with said first recombinant viral vector then co-infected with said second recombinant plasmid such that the DNA molecule encoding the HCV/SEAP reporter gene is under control of said promoter; and
- d) the target mammalian cell expresses said HCV/SEAP reporter gene polyprotein such that SEAP is secreted from said target mammalian

cell.

37. The isolated DNA sequence of claim 27 comprising a DNA sequence or variants thereof in SEQ. ID NO. 8.
38. The isolated DNA sequence of claim 28 comprising a DNA sequence or variants thereof in SEQ. ID NO. 15.
39. A composition comprising the pHCAP1 polyprotein as described in SEQ. ID NO. 2.
40. A composition comprising the pHCAP3 polyprotein as described in SEQ. ID NO. 9.
41. A composition comprising the pHCAP4 polyprotein as described in SEQ. ID NO. 16.

## Vaccinia Virus NS3/SEAP System

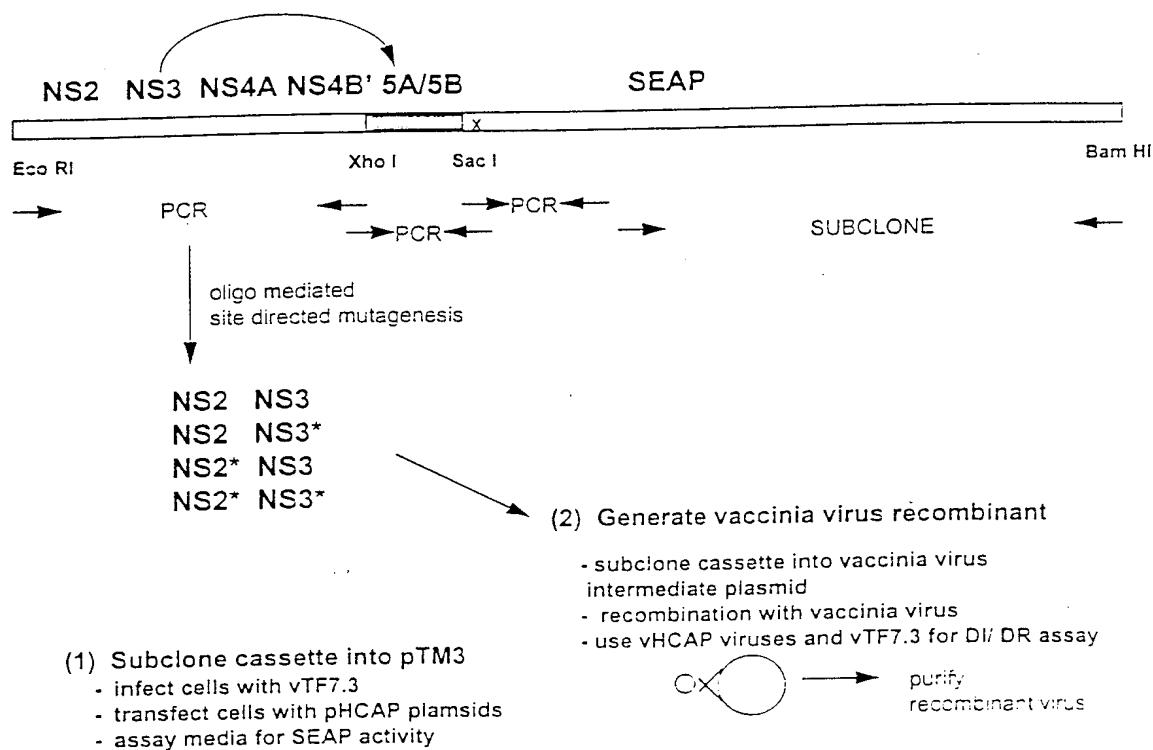


Figure 1

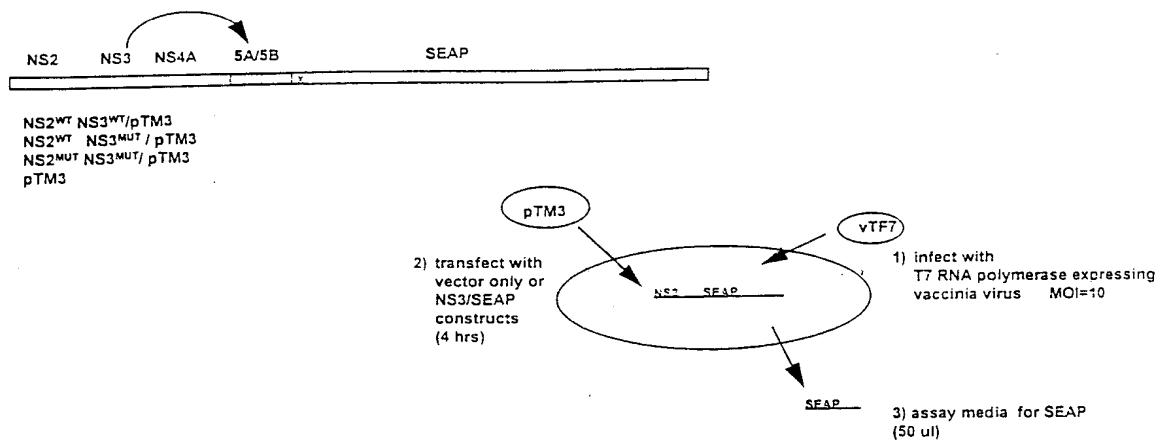


Figure 1B

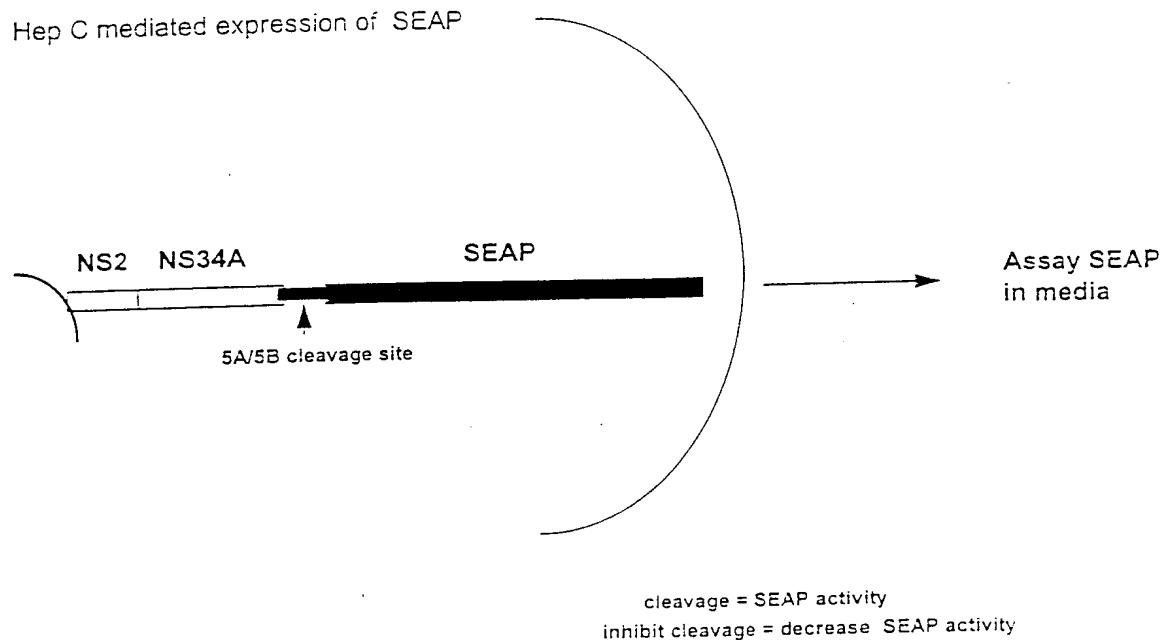


Figure 2

## DI/DR Assay

vTF7.3 (T7 RNA polymerase recombinant)  
 vICAP1 (ICAP1/ICAP3/ICAP recombinant)  
 vICAP3 (ICAP1/ICAP3/ICAP recombinant)

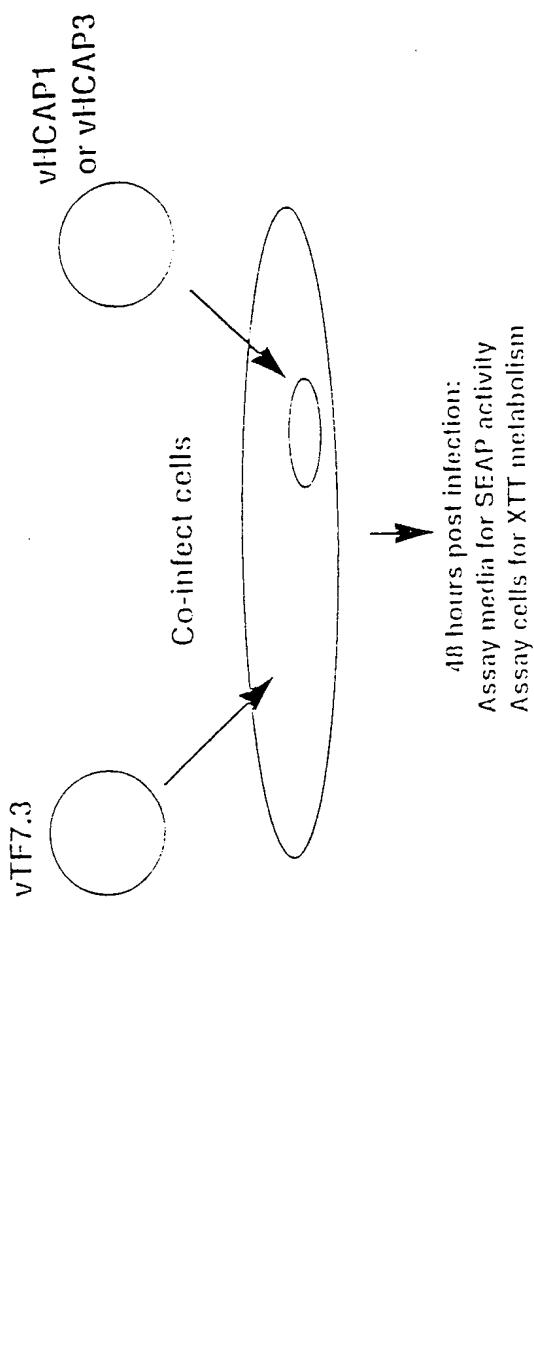


Figure 3

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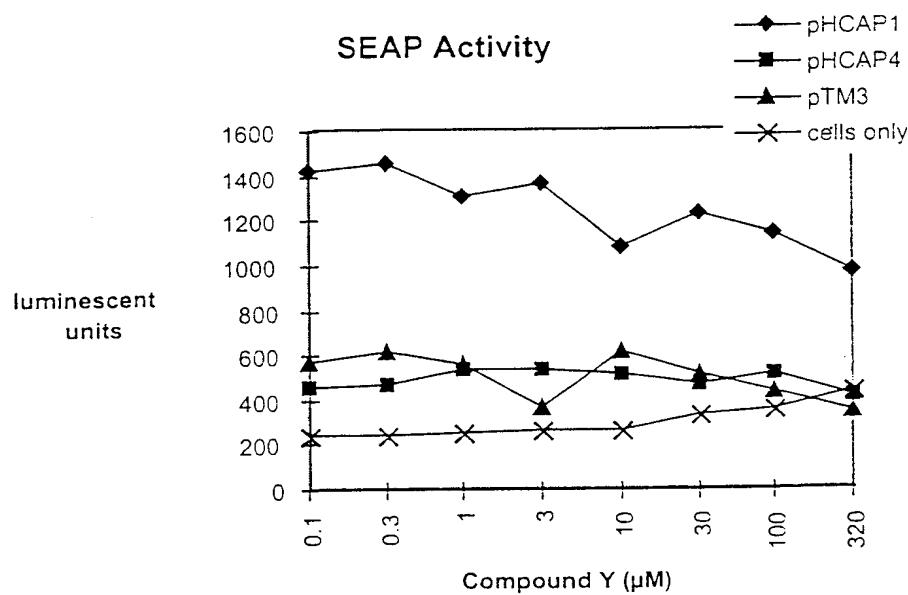
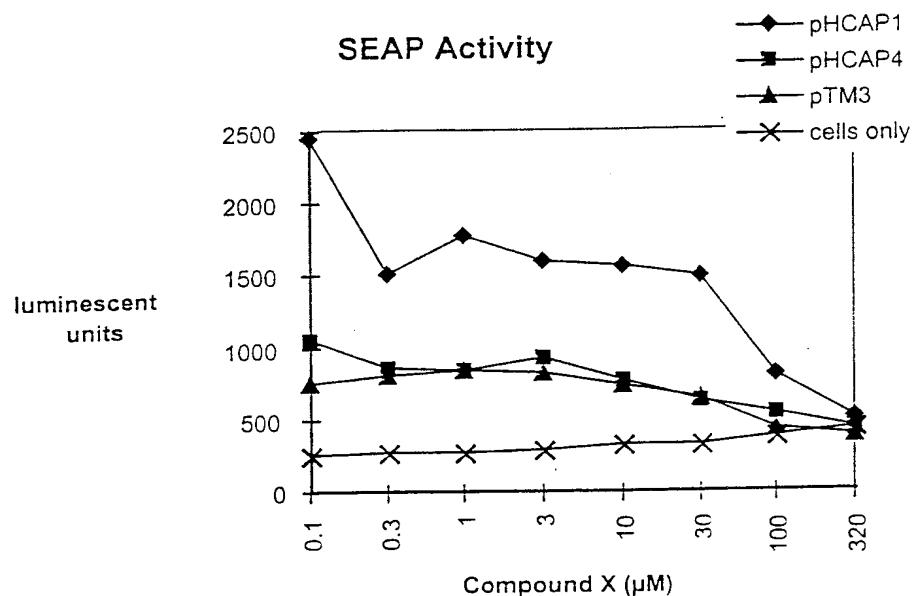


Figure 4 A

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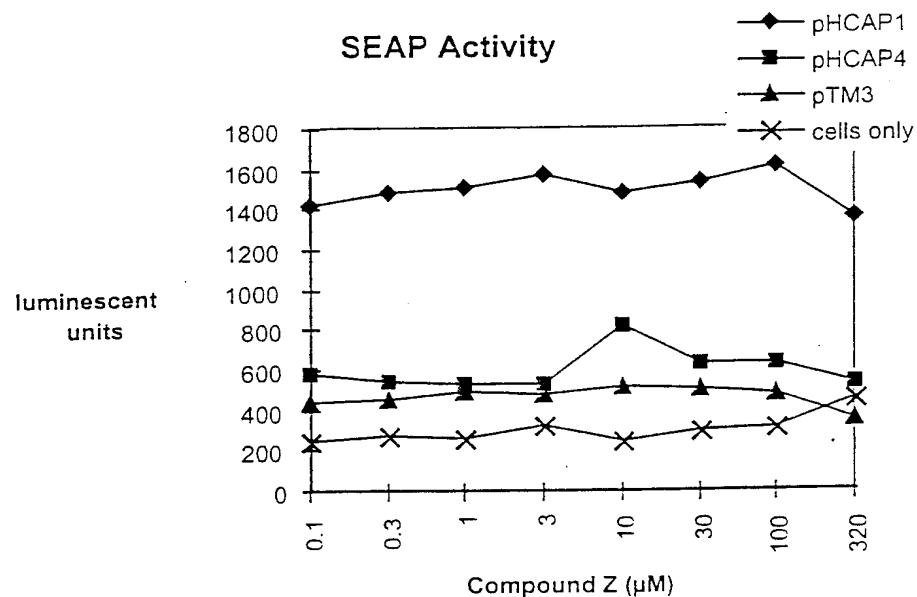


Figure 4 B

Figure 5

Figure 6.

Figure 7

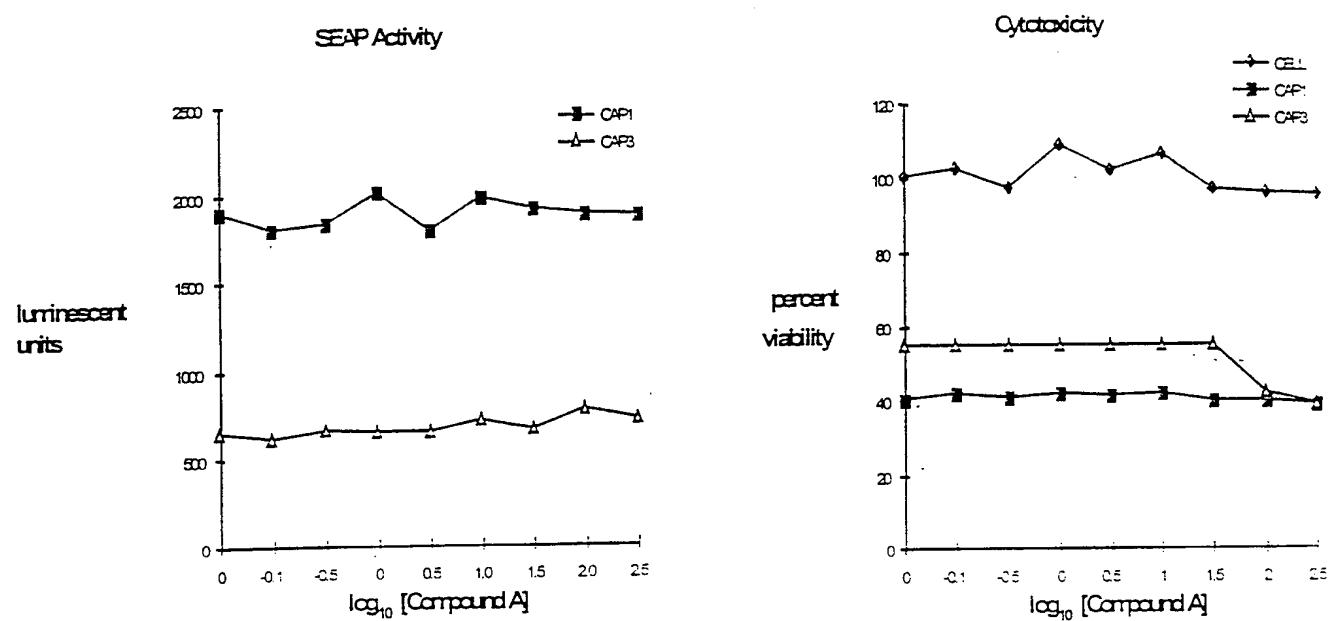


Figure 8

## DI/ DR Assay Compound Summary

Compound	EC <sub>50</sub> (μM)	CC <sub>50</sub> (μM)	TI	Solubility	Activity
A	> 320	> 320	-	> 320	-
B	18	15	1	> 320	-
C	37	41	1	> 320	-
D	> 320	> 320	-	> 320	-
E	70	174	2	ppt > 30	-
F	64	> 320	4	> 320	+/-
G	> 320	> 320	-	> 320	-
H	166	194	1	> 320	-
I	38	76	2	> 320	-

Platemap:

11 / 11

	1	2	3	4	5	6	7	8	9	10	11	12	Compounds
Compound	μM	0•	0•	320•	100•	30•	10•	3•	1•	0.3•	0.1•	0•	0•
A	BHK ONLY												
B	pHCAP1												
C	pHCAP1												
D	pHCAP4												
E	pHCAP4												
F	pTM3												
G	pTM3												
H	BHK ONLY												

W  
X  
Y  
Z

Figure 9

## SEQUENCE LISTING

<110> Potts, Karen E.  
Jackson, Roberta L.  
Patick, Amy K.

<120> REPORTER GENE SYSTEM FOR USE IN CELL-BASED ASSESSMENT  
OF INHIBITORS OF THE HEPATITIS C VIRUS PROTEASE

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 705 710 715 720  
 Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn Ile Glu Glu Val  
 725 730 735  
 Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly Lys Ala Ile Pro  
 740 745 750  
 Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe Cys His Ser Lys  
 755 760 765  
 Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly Leu Gly Ile Asn  
 770 775 780  
 Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Val Ile Pro Thr Ile  
 785 790 795 800  
 Gly Asp Val Val Val Ala Thr Asp Ala Leu Met Thr Gly Tyr Thr  
 805 810 815  
 Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys Val Thr Gln Thr  
 820 825 830  
 Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu Thr Thr Thr Val  
 835 840 845  
 Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly Arg Thr Gly Arg  
 850 855 860  
 Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly Glu Arg Pro Ser  
 865 870 875 880  
 Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr Asp Ala Gly Cys  
 885 890 895  
 Ala Trp Tyr Glu Leu Thr Pro Ala Glu Thr Ser Val Arg Leu Arg Ala  
 900 905 910  
 Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp His Leu Glu Phe  
 915 920 925  
 Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp Ala His Phe Leu  
 930 935 940  
 Ser Gln Thr Lys Gln Ala Gly Asp Asn Phe Pro Tyr Leu Val Ala Tyr  
 945 950 955 960  
 Gln Ala Thr Val Cys Ala Arg Ala Gln Ala Pro Pro Pro Ser Trp Asp  
 965 970 975

Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr Leu His Gly Pro  
 980 985 990  
 Thr Pro Leu Leu Tyr Arg Leu Gly Ala Val Gln Asn Glu Val Thr Leu  
 995 1000 1005  
 Thr His Pro Ile Thr Lys Tyr Ile Met Ala Cys Met Ser Ala Asp Leu  
 1010 1015 1020  
 Glu Val Val Thr Ser Thr Trp Val Leu Val Gly Gly Val Leu Ala Ala  
 025 1030 1035 1040  
 Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val Val Ile Val Gly Arg  
 1045 1050 1055  
 Ile Ile Leu Ser Gly Arg Pro Ala Ile Val Pro Asp Arg Glu Leu Leu  
 1060 1065 1070  
 Tyr Gln Glu Phe Asp Glu Met Glu Glu Cys Ala Ser His Leu Pro Tyr  
 1075 1080 1085  
 Ile Glu Gln Gly Met Gln Leu Ala Glu Gln Phe Lys Gln Lys Ala Leu  
 1090 1095 1100  
 Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala Ala Ala Pro Val  
 105 1110 1115 1120  
 Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp Ala Lys His Met  
 1125 1130 1135  
 Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly Leu Ser Thr Leu  
 1140 1145 1150  
 Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe Thr Ala Ser Ile  
 1155 1160 1165  
 Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe Asn Ile Leu Gly  
 1170 1175 1180  
 Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser Ala Ala Ser Ala Phe  
 185 1190 1195 1200  
 Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly Ser Ile Gly Leu Gly  
 1205 1210 1215  
 Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala Gly Val Ala Gly  
 1220 1225 1230  
 Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met Pro Ser Thr Glu  
 1235 1240 1245  
 Asp Leu Val Asn Leu Leu Pro Ala Ile Leu Glu Glu Ala Ser Glu Asp  
 1250 1255 1260  
 Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly Ala Leu Glu Leu  
 265 1270 1275 1280  
 Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu Ser Leu Gly Ile  
 1285 1290 1295

Ile Pro Val Glu Glu Glu Asn Pro Asp Phe Trp Asn Arg Glu Ala Ala  
 1300 1305 1310

Glu Ala Leu Gly Ala Ala Lys Lys Leu Gln Pro Ala Gln Thr Ala Ala  
 1315 1320 1325

Lys Asn Leu Ile Ile Phe Leu Gly Asp Gly Met Gly Val Ser Thr Val  
 1330 1335 1340

Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp Lys Leu Gly Pro  
 345 1350 1355 1360

Glu Ile Pro Leu Ala Met Asp Arg Phe Pro Tyr Val Ala Leu Ser Lys  
 1365 1370 1375

Thr Tyr Asn Val Asp Lys His Val Pro Asp Ser Gly Ala Thr Ala Thr  
 1380 1385 1390

Ala Tyr Leu Cys Gly Val Lys Gly Asn Phe Gln Thr Ile Gly Leu Ser  
 1395 1400 1405

Ala Ala Ala Arg Phe Asn Gln Cys Asn Thr Thr Arg Gly Asn Glu Val  
 1410 1415 1420

Ile Ser Val Met Asn Arg Ala Lys Lys Ala Gly Lys Ser Val Gly Val  
 425 1430 1435 1440

Val Thr Thr Arg Val Gln His Ala Ser Pro Ala Gly Thr Tyr Ala  
 1445 1450 1455

His Thr Val Asn Arg Asn Trp Tyr Ser Asp Ala Asp Val Pro Ala Ser  
 1460 1465 1470

Ala Arg Gln Glu Gly Cys Gln Asp Ile Ala Thr Gln Leu Ile Ser Asn  
 1475 1480 1485

Met Asp Ile Asp Val Ile Leu Gly Gly Arg Lys Tyr Met Phe Pro  
 1490 1495 1500

Met Gly Thr Pro Asp Pro Glu Tyr Pro Asp Asp Tyr Ser Gln Gly Gly  
 505 1510 1515 1520

Thr Arg Leu Asp Gly Lys Asn Leu Val Gln Glu Trp Leu Ala Lys Arg  
 1525 1530 1535

Gln Gly Ala Arg Tyr Val Trp Asn Arg Thr Glu Leu Met Gln Ala Ser  
 1540 1545 1550

Leu Asp Pro Ser Val Thr His Leu Met Gly Leu Phe Glu Pro Gly Asp  
 1555 1560 1565

Met Lys Tyr Glu Ile His Arg Asp Ser Thr Leu Asp Pro Ser Leu Met  
 1570 1575 1580

Glu Met Thr Glu Ala Ala Leu Arg Leu Leu Ser Arg Asn Pro Arg Gly  
 585 1590 1595 1600

Phe Phe Leu Phe Val Glu Gly Gly Arg Ile Asp His Gly His His Glu  
 1605 1610 1615

Ser Arg Ala Tyr Arg Ala Leu Thr Glu Thr Ile Met Phe Asp Asp Ala  
 1620 1625 1630  
 Ile Glu Arg Ala Gly Gln Leu Thr Ser Glu Glu Asp Thr Leu Ser Leu  
 1635 1640 1645  
 Val Thr Ala Asp His Ser His Val Phe Ser Phe Gly Gly Tyr Pro Leu  
 1650 1655 1660  
 Arg Gly Ser Cys Ile Phe Gly Leu Ala Pro Gly Lys Ala Arg Asp Arg  
 665 1670 1675 1680  
 Lys Ala Tyr Thr Val Leu Leu Tyr Gly Asn Gly Pro Gly Tyr Val Leu  
 1685 1690 1695  
 Lys Asp Gly Ala Arg Pro Asp Val Thr Glu Ser Glu Ser Gly Ser Pro  
 1700 1705 1710  
 Glu Tyr Arg Gln Gln Ser Ala Val Pro Leu Asp Glu Glu Thr His Ala  
 1715 1720 1725  
 Gly Glu Asp Val Ala Val Phe Ala Arg Gly Pro Gln Ala His Leu Val  
 1730 1735 1740  
 His Gly Val Gln Glu Gln Thr Phe Ile Ala His Val Met Ala Phe Ala  
 745 1750 1755 1760  
 Ala Cys Leu Glu Pro Tyr Thr Ala Cys Asp Leu Ala Pro Pro Ala Gly  
 1765 1770 1775  
 Thr Thr Asp Ala Ala His Pro Gly Met Ser Glu Lys Tyr Ile Val Thr  
 1780 785 1790  
 Trp Asp Met Leu Gln Ile His Ala Arg Lys Leu Ala Ser Arg Leu Met  
 1795 1800 1805  
 Pro Ser Glu Gln Trp Lys Gly Ile Ile Ala Val Ser Arg Gly Gly Leu  
 1810 1815 1820  
 Val Pro Gly Ala Leu Leu Ala Arg Glu Leu Gly Ile Arg His Val Asp  
 825 1830 1835 1840  
 Thr Val Cys Ile Ser Ser Tyr Asp His Asp Asn Gln Arg Glu Leu Lys  
 1845 1850 1855  
 Val Leu Lys Arg Ala Glu Gly Asp Gly Glu Gly Phe Ile Val Ile Asp  
 1860 1865 1870  
 Asp Leu Val Asp Thr Gly Gly Thr Ala Val Ala Ile Arg Glu Met Tyr  
 1875 1880 1885  
 Pro Lys Ala His Phe Val Thr Ile Phe Ala Lys Pro Ala Gly Arg Pro  
 1890 1895 1900  
 Leu Val Asp Asp Tyr Val Val Asp Ile Pro Gln Asp Thr Trp Ile Glu  
 905 1910 1915 1920  
 Gln Pro Trp Asp Met Gly Val Val Phe Val Pro Pro Ile Ser Gly Arg  
 1925 1930 1935

Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val Ala Ala  
 1940 1945 1950

Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu Asn Leu  
 1955 1960 1965

Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys Met Lys  
 1970 1975 1980

Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Gly Glu Glu Thr Glu  
 985 1990 1995 2000

Ile Glu Ile Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys Arg Lys  
 2005 2010 2015

Cys Tyr Ile Asp Ser Met Ser Ile Gln His Phe Arg Val Ala Leu Ile  
 2020 2025 2030

Pro Phe Phe Ala Ala Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr  
 2035 2040 2045

Leu Val Lys Val Lys Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly  
 2050 2055 2060

Tyr Ile Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg  
 065 2070 2075 208

Pro Glu Glu Arg Phe Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys  
 2085 2090 2095

Gly Ala Val Leu Ser Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly Arg  
 2100 2105 2110

Arg Ile His Tyr Ser Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr  
 2115 2120 2125

Glu Lys His Leu Thr Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala  
 2130 2135 2140

Ala Ile Thr Met Ser Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr  
 145 2150 2155 216

Ile Gly Gly Pro Lys Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp  
 2165 2170 2175

His Val Thr Arg Leu Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile  
 2180 2185 2190

Pro Asn Asp Glu Arg Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr  
 2195 2200 2205

Leu Arg Lys Leu Leu Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln  
 2210 2215 2220

Gln Leu Ile Asp Trp Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu  
 225 2230 2235 224

Arg Ser Ala Leu Pro Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala  
 2245 2250 2255

Gly Glu Arg Gly Ser Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly  
 2260 2265 2270

Lys Pro Ser Arg Ile Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr  
 2275 2280 2285

Met Asp Glu Arg Asn Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile  
 2290 2295 2300

Lys His Trp  
 305

<210> 3  
 <211> 92  
 <212> PRT  
 <213> Artificial Sequence

<400> 3  
 Met Asn Gly Gly His Ile Gln Leu Ile Ile Gly Pro Met Phe Ser Gly  
 1 5 10 15

Lys Ser Thr Glu Leu Ile Arg Arg Val Arg Arg Tyr Gln Ile Ala Gln  
 20 25 30

Tyr Lys Cys Val Thr Ile Lys Tyr Ser Asn Asp Asn Arg Tyr Gly Thr  
 35 40 45

Gly Leu Trp Thr His Asp Lys Asn Asn Phe Glu Ala Leu Glu Ala Thr  
 50 55 60

Lys Leu Cys Asp Val Leu Glu Ser Ile Thr Asp Phe Ser Val Ile Gly  
 65 70 75 80

Ile Asp Glu Gly Gln Phe Phe Pro Asp Ile Val Glu  
 85 90

<210> 4  
 <211> 1692  
 <212> PRT  
 <213> Artificial Sequence

<400> 4  
 Met Gly Ile Pro Gln Phe Met Ala Arg Val Cys Ala Cys Leu Trp Met  
 1 5 10 15

Met Leu Leu Ile Ala Gln Ala Glu Ala Ala Leu Glu Asn Leu Val Val  
 20 25 30

Leu Asn Ala Ala Ser Val Ala Gly Ala His Gly Ile Leu Ser Phe Leu  
 35 40 45

Val Phe Phe Cys Ala Ala Trp Tyr Ile Lys Gly Arg Leu Val Pro Gly  
 50 55 60

Ala Ala Tyr Ala Leu Tyr Gly Val Trp Pro Leu Leu Leu Leu Leu  
 65 70 75 80

Ala Leu Pro Pro Arg Ala Tyr Ala Met Asp Arg Glu Met Ala Ala Ser  
 85 90 95

Cys Gly Gly Ala Val Phe Val Gly Leu Val Leu Leu Thr Leu Ser Pro  
 100 105 110  
 Tyr Tyr Lys Val Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr Phe  
 115 120 125  
 Thr Thr Arg Ala Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn  
 130 135 140  
 Ala Arg Gly Gly Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His  
 145 150 155 160  
 Pro Glu Leu Ile Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly  
 165 170 175  
 Pro Leu Met Val Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val  
 180 185 190  
 Arg Ala Gln Gly Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala  
 195 200 205  
 Gly Gly His Tyr Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr  
 210 215 220  
 Gly Thr Tyr Ile Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His  
 225 230 235 240  
 Ala Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser  
 245 250 255  
 Asp Met Glu Thr Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala Cys  
 260 265 270  
 Gly Asp Ile Ile Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu  
 275 280 285  
 Ile Leu Leu Gly Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu  
 290 295 300  
 Leu Ala Pro Ile Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly  
 305 310 315 320  
 Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly  
 325 330 335  
 Glu Val Gln Val Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys  
 340 345 350  
 Val Asn Gly Val Cys Trp Thr Val Tyr His Gly Ala Gly Ser Lys Thr  
 355 360 365  
 Leu Ala Gly Pro Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp  
 370 375 380  
 Gln Asp Leu Val Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr  
 385 390 395 400  
 Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala  
 405 410 415

Asp Val Ile Pro Val Arg Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu  
 420 425 430

Ser Pro Arg Pro Val Ser Tyr Leu Lys Gly Ser Ser Gly Gly Pro Leu  
 435 440 445

Leu Cys Pro Ser Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys  
 450 455 460

Thr Arg Gly Val Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met  
 465 470 475 480

Glu Thr Thr Met Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro  
 485 490 495

Ala Val Pro Gln Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly  
 500 505 510

Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr  
 515 520 525

Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly  
 530 535 540

Ala Tyr Met Ser Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly  
 545 550 555 560

Val Arg Thr Ile Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly  
 565 570 575

Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile  
 580 585 590

Ile Cys Asp Glu Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile  
 595 600 605

Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val  
 610 615 620

Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn  
 625 630 635 640

Ile Glu Glu Val Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly  
 645 650 655

Lys Ala Ile Pro Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe  
 660 665 670

Cys His Ser Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly  
 675 680 685

Leu Gly Ile Asn Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Val  
 690 695 700

Ile Pro Thr Ile Gly Asp Val Val Val Ala Thr Asp Ala Leu Met  
 705 710 715 720

Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys  
 725 730 735

Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu  
 740 745 750  
 Thr Thr Thr Val Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly  
 755 760 765  
 Arg Thr Gly Arg Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly  
 770 775 780  
 Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr  
 785 790 795 800  
 Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro Ala Glu Thr Ser Val  
 805 810 815  
 Arg Leu Arg Ala Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp  
 820 825 830  
 His Leu Glu Phe Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp  
 835 840 845  
 Ala His Phe Leu Ser Gln Thr Lys Gln Ala Gly Asp Asn Phe Pro Tyr  
 850 855 860  
 Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Gln Ala Pro Pro  
 865 870 875 880  
 Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr  
 885 890 895  
 Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ala Val Gln Asn  
 900 905 910  
 Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr Ile Met Ala Cys Met  
 915 920 925  
 Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp Val Leu Val Gly Gly  
 930 935 940  
 Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val Val  
 945 950 955 960  
 Ile Val Gly Arg Ile Ile Leu Ser Gly Arg Pro Ala Ile Val Pro Asp  
 965 970 975  
 Arg Glu Leu Leu Tyr Gln Glu Phe Asp Glu Met Glu Glu Cys Ala Ser  
 980 985 990  
 His Leu Pro Tyr Ile Glu Gln Gly Met Gln Leu Ala Glu Gln Phe Lys  
 995 1000 1005  
 Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala  
 1010 1015 1020  
 Ala Ala Pro Val Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp  
 1025 1030 1035 1040  
 Ala Lys His Met Trp Asn Phe Ser Gly Ile Gln Tyr Leu Ala Gly  
 1045 1050 1055

Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe  
 1060 1065 1070  
 Thr Ala Ser Ile Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe  
 1075 1080 1085  
 Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser Ala  
 1090 1095 1100  
 Ala Ser Ala Phe Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly Ser  
 105 1110 1115 1120  
 Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala  
 1125 1130 1135  
 Gly Val Ala Gly Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met  
 1140 1145 1150  
 Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro Ala Ile Leu Glu Glu  
 1155 1160 1165  
 Ala Ser Glu Asp Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly  
 1170 1175 1180  
 Ala Leu Glu Leu Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu  
 185 1190 1195 1200  
 Ser Leu Gly Ile Ile Pro Val Glu Glu Asn Pro Asp Phe Trp Asn  
 1205 1210 1215  
 Arg Glu Ala Ala Glu Ala Leu Gly Ala Ala Lys Lys Leu Gln Pro Ala  
 1220 1225 1230  
 Gln Thr Ala Ala Lys Asn Leu Ile Ile Phe Leu Gly Asp Gly Met Gly  
 1235 1240 1245  
 Val Ser Thr Val Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp  
 1250 1255 1260  
 Lys Leu Gly Pro Glu Ile Pro Leu Ala Met Asp Arg Phe Pro Tyr Val  
 265 1270 1275 1280  
 Ala Leu Ser Lys Thr Tyr Asn Val Asp Lys His Val Pro Asp Ser Gly  
 1285 1290 1295  
 Ala Thr Ala Thr Ala Tyr Leu Cys Gly Val Lys Gly Asn Phe Gln Thr  
 1300 1305 1310  
 Ile Gly Leu Ser Ala Ala Ala Arg Phe Asn Gln Cys Asn Thr Thr Arg  
 1315 1320 1325  
 Gly Asn Glu Val Ile Ser Val Met Asn Arg Ala Lys Lys Ala Gly Lys  
 1330 1335 1340  
 Ser Val Gly Val Val Thr Thr Arg Val Gln His Ala Ser Pro Ala  
 345 1350 1355 1360  
 Gly Thr Tyr Ala His Thr Val Asn Arg Asn Trp Tyr Ser Asp Ala Asp  
 1365 1370 1375

Val Pro Ala Ser Ala Arg Gln Glu Gly Cys Gln Asp Ile Ala Thr Gln  
 1380 1385 1390  
 Leu Ile Ser Asn Met Asp Ile Asp Val Ile Leu Gly Gly Gly Arg Lys  
 1395 1400 1405  
 Tyr Met Phe Pro Met Gly Thr Pro Asp Pro Glu Tyr Pro Asp Asp Tyr  
 1410 1415 1420  
 Ser Gln Gly Gly Thr Arg Leu Asp Gly Lys Asn Leu Val Gln Glu Trp  
 425 1430 1435 1440  
 Leu Ala Lys Arg Gln Gly Ala Arg Tyr Val Trp Asn Arg Thr Glu Leu  
 1445 1450 1455  
 Met Gln Ala Ser Leu Asp Pro Ser Val Thr His Leu Met Gly Leu Phe  
 1460 1465 1470  
 Glu Pro Gly Asp Met Lys Tyr Glu Ile His Arg Asp Ser Thr Leu Asp  
 1475 1480 1485  
 Pro Ser Leu Met Glu Met Thr Glu Ala Ala Leu Arg Leu Leu Ser Arg  
 1490 1495 1500  
 Asn Pro Arg Gly Phe Phe Leu Phe Val Glu Gly Gly Arg Ile Asp His  
 505 1510 1515 1520  
 Gly His His Glu Ser Arg Ala Tyr Arg Ala Leu Thr Glu Thr Ile Met  
 1525 1530 1535  
 Phe Asp Asp Ala Ile Glu Arg Ala Gly Gln Leu Thr Ser Glu Glu Asp  
 1540 1545 1550  
 Thr Leu Ser Leu Val Thr Ala Asp His Ser His Val Phe Ser Phe Gly  
 1555 1560 1565  
 Gly Tyr Pro Leu Arg Gly Ser Cys Ile Phe Gly Leu Ala Pro Gly Lys  
 1570 1575 1580  
 Ala Arg Asp Arg Lys Ala Tyr Thr Val Leu Leu Tyr Gly Asn Gly Pro  
 585 1590 1595 1600  
 Gly Tyr Val Leu Lys Asp Gly Ala Arg Pro Asp Val Thr Glu Ser Glu  
 1605 1610 1615  
 Ser Gly Ser Pro Glu Tyr Arg Gln Gln Ser Ala Val Pro Leu Asp Glu  
 1620 1625 1630  
 Glu Thr His Ala Gly Glu Asp Val Ala Val Phe Ala Arg Gly Pro Gln  
 1635 1640 1645  
 Ala His Leu Val His Gly Val Gln Glu Gln Thr Phe Ile Ala His Val  
 1650 1655 1660  
 Met Ala Phe Ala Ala Cys Leu Glu Pro Tyr Thr Ala Cys Asp Leu Ala  
 665 1670 1675 1680  
 Pro Pro Ala Gly Thr Thr Asp Ala Ala His Pro Gly  
 1685 1690

<210> 5  
<211> 152  
<212> PRT  
<213> Artificial Sequence

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Met Ser Glu Lys Tyr Ile Val Thr Trp Asp Met Leu Gln Ile His Ala  
1 5 10 15  
Arg Lys Leu Ala Ser Arg Leu Met Pro Ser Glu Gln Trp Lys Gly Ile  
20 25 30  
Ile Ala Val Ser Arg Gly Gly Leu Val Pro Gly Ala Leu Leu Ala Arg  
35 40 45  
Glu Leu Gly Ile Arg His Val Asp Thr Val Cys Ile Ser Ser Tyr Asp  
50 55 60  
His Asp Asn Gln Arg Glu Leu Lys Val Leu Lys Arg Ala Glu Gly Asp  
65 70 75 80  
Gly Glu Gly Phe Ile Val Ile Asp Asp Leu Val Asp Thr Gly Gly Thr  
85 90 95  
Ala Val Ala Ile Arg Glu Met Tyr Pro Lys Ala His Phe Val Thr Ile  
100 105 110  
Phe Ala Lys Pro Ala Gly Arg Pro Leu Val Asp Asp Tyr Val Val Asp  
115 120 125  
Ile Pro Gln Asp Thr Trp Ile Glu Gln Pro Trp Asp Met Gly Val Val  
130 135 140  
Phe Val Pro Pro Ile Ser Gly Arg  
145 150

<210> 6  
<211> 85  
<212> PRT  
<213> Artificial Sequence

<400> 6  
Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val Ala Ala  
1 5 10 15  
Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu Asn Leu  
20 25 30  
Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys Met Lys  
35 40 45  
Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Gly Glu Glu Thr Glu  
50 55 60  
Ile Glu Ile Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys Arg Lys  
65 70 75 80  
Cys Tyr Ile Asp Ser  
85

<210> 7  
<211> 286  
<212> PRT  
<213> Artificial Sequence

<400> 7  
Met Ser Ile Gln His Phe Arg Val Ala Leu Ile Pro Phe Phe Ala Ala  
1 5 10 15  
Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr Leu Val Lys Val Lys  
20 25 30  
Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu Leu Asp  
35 40 45  
Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg Pro Glu Glu Arg Phe  
50 55 60  
Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala Val Leu Ser  
65 70 75 80  
Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly Arg Arg Ile His Tyr Ser  
85 90 95  
Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His Leu Thr  
100 105 110  
Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala Ala Ile Thr Met Ser  
115 120 125  
Asp Asn Thr Ala Ala Asn Leu Leu Thr Thr Ile Gly Gly Pro Lys  
130 135 140  
Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp His Val Thr Arg Leu  
145 150 155 160  
Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg  
165 170 175  
Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr Leu Arg Lys Leu Leu  
180 185 190  
Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp  
195 200 205  
Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu Arg Ser Ala Leu Pro  
210 215 220  
Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly Glu Arg Gly Ser  
225 230 235 240  
Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile  
245 250 255  
Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr Met Asp Glu Arg Asn  
260 265 270  
Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile Lys His Trp  
275 280 285

<210> 8  
<211> 13910  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: plasmid phcap 3

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<221> CDS  
<222> (497) .. (772)

<220>  
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<222> (8579) .. (9034)

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<222> (10191) .. (10445)

<220>  
<221> CDS  
<222> (11877) .. (12734)

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<221> misc\_feature  
<222> (1) .. (774)  
<223> Vaccinia Virus thymidine Kinase gene recombination site

<220>  
<221> promoter  
<222> (794) .. (816)  
<223> T7 promoter

<220>  
<221> misc\_feature  
<222> (846) .. (1424)  
<223> EMC/Internal Ribosome Entry Site (IRES)

<220>  
<221> misc\_feature  
<222> (1426) .. (1437)  
<223> MCS (Multiple Cloning Site)

<220>  
<221> misc\_feature  
<222> (1446) .. (2318)  
<223> HCV E2/ NS2 domain

<220>  
<221> misc\_feature  
<222> (2319) .. (4231)  
<223> HCV NS3 Domain containing the serine protease and helicase enzymes

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 aatggcagaa attcgccgga tctttgtgaa ggaaccttac ttctgtggtg tgacataatt 7060  
 ggacaaaacta cctacagaga tttaaagctc taaggtaaat ataaaatttt taagtgtata 7120  
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 caggctggga cacttcac atg agc gaa aaa tac atc gtc acc tgg gac atg 8611  
                   Met Ser Glu Lys Tyr Ile Val Thr Trp Asp Met  
                   1785               1790               1795  
 ttg cag atc cat gca cgt aaa ctc gca agc cga ctg atg cct tct gaa 8659  
 Leu Gln Ile His Ala Arg Lys Leu Ala Ser Arg Leu Met Pro Ser Glu  
                   1800               1805               1810  
 caa tgg aaa ggc att att gcc gta agc cgt ggc ggt ctg gta ccg ggt 8707  
 Gln Trp Lys Gly Ile Ala Val Ser Arg Gly Gly Leu Val Pro Gly  
                   1815               1820               1825  
 gcg tta ctg gcg cgt gaa ctg ggt att cgt cat gtc gat acc gtt tgt 8755  
 Ala Leu Leu Ala Arg Glu Leu Gly Ile Arg His Val Asp Thr Val Cys  
                   1830               1835               1840  
 att tcc agc tac gat cac gac aac cag cgc gag ctt aaa gtg ctg aaa 8803  
 Ile Ser Ser Tyr Asp His Asp Asn Gln Arg Glu Leu Lys Val Leu Lys  
                   1845               1850               1855  
 cgc gca gaa ggc gat ggc gaa ggc ttc atc gtt att gat gac ctg gtg 8851  
 Arg Ala Glu Gly Asp Gly Glu Gly Phe Ile Val Ile Asp Asp Leu Val  
                   1860               1865               1870               1875  
 gat acc ggt ggt act gcg gtt gcg att cgt gaa atg tat cca aaa gcg 8899  
 Asp Thr Gly Gly Thr Ala Val Ala Ile Arg Glu Met Tyr Pro Lys Ala  
                   1880               1885               1890  
 cac ttt gtc acc atc ttc gca aaa ccg gct ggt cgt ccg ctg gtt gat 8947  
 His Phe Val Thr Ile Phe Ala Lys Pro Ala Gly Arg Pro Leu Val Asp  
                   1895               1900               1905  
 gac tat gtt gtt gat atc ccg caa gat acc tgg att gaa cag ccg tgg 8995  
 Asp Tyr Val Val Asp Ile Pro Gln Asp Thr Trp Ile Glu Gln Pro Trp  
                   1910               1915               1920  
 gat atg ggc gtc gta ttc gtc ccg cca atc tcc ggt cgc taatctttc 9044  
 Asp Met Gly Val Val Phe Val Pro Pro Ile Ser Gly Arg  
                   1925               1930               1935  
 aacgcctggc actgccgggc gttgttctt ttaacttcag gcgggttaca atagttcca 9104  
 gtaagtattc tggaggctgc atccatgaca caggcaaacc tgagcgaaac cctgttcaaa 9164  
 ccccgcttta aacatcctga aacctcgacg ctagtccgcc gctttaatca cggcgacaaa 9224  
 ccgcctgtgc agtcggccct tcatggtaaa accatccctc actggtatcg catgattaac 9284  
 cgtctgatgt ggatctggcg cggcattgac ccacgcgaaa tcctcgacgt ccaggcacgt 9344

attgtatga gcgatgccga acgtaccgac gatgatttat acgatacggt gattggctac 9404  
 cgtggccgca actggattta tgagtggcc ccggatctt gtgaaggaac cttacttctg 9464  
 tggtgaca taattggaca aactacctac agagattaa agctctaagg taaatataaa 9524  
 attttaagt gtataatgtg taaaactact gattctaatt gtttgtgtat ttttagattcc 9584  
 aacctatgga actgatgaat gggagcagtg gtggaatgcc tttatgagg aaaacctgtt 9644  
 ttgctcagaa gaaatgccat ctagtgatga tgaggctact gctgactctc aacattctac 9704  
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 aagtaggcat aacagttata atcataacat actgtttttt cttaactccac acaggcatag 9944  
 agtgtctgct attaataact atgctcaaaa attgtgtacc tttagcttt taatttgtaa 10004  
 aggggttaat aaggaatatt tgatgtatacg tgccttgact agagatcata atcagccata 10064  
 ccacatttgt agaggtttta cttgctttaaaaacccccc acacccccc ctgaacctga 10124  
 aacataaaat gaatgcaatt gttgttgtt aagttttttt aattgcatgc tccggatcga 10184  
 gatcaa ttc tgt gag cgt atg gca aac gaa gga aaa ata gtt ata gta 10232  
     Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val  
     1940                       1945                       1950  
 gcc gca ctc gat ggg aca ttt caa cgt aaa ccg ttt aat aat att ttg 10280  
     Ala Ala Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu  
     1955                       1960                       1965  
 aat ctt att cca tta tct gaa atg gtg gta aaa cta act gct gtg tgt 10328  
     Asn Leu Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys  
     1970                       1975                       1980  
 atg aaa tgc ttt aag gag gct tcc ttt tct aaa cga ttg ggt gag gaa 10376  
     Met Lys Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Glu Glu  
     1985                       1990                       1995  
 acc gag ata gaa ata ata gga ggt aat gat atg tat caa tcg gtg tgt 10424  
     Thr Glu Ile Glu Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys  
     2000                       2005                       2010  
 aga aag tgt tac atc gac tca taatattata ttttttatct aaaaaactaa 10475  
     Arg Lys Cys Tyr Ile Asp Ser  
     2015                       2020  
 aaataaaacat tgattaaatt ttaatataat actaaaaat ggatgttgtg tcgttagata 10535  
 aaccgtttat gtatTTTgag gaaattgata atgagttaga ttacgaacca gaaagtgc 10595  
 atgaggtcgc aaaaaaactg ccgtatcaag gacagttaaa actattacta ggagaattat 10655  
 tttttcttag taagttacag cgacacggta tattagatgg tgccaccgta gtgtatata 10715  
 gatctgctcc cggtacacat atacgttatt tgagagatca tttctataat ttaggagtga 10775

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atgtgactct agtgactcgg ttctgttgcg aggaatatct acgatccatc aaaaaacaac 10895  
tgcatttttc taagattattt ttaatttctg atgtgagatc caaacgagga gaaaatgaac 10955  
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ttctaaatac attcaaataat gtatccgctc atgagacaat aaccctgata aatgcttcaa 11855  
taatattgaa aaaggaagag t atg agt att caa cat ttc cgt gtc gcc ctt 11906  
Met Ser Ile Gln His Phe Arg Val Ala Leu  
2025 2030  
  
att ccc ttt ttt gcg gca ttt tgc ctt cct gtt ttt gct cac cca gaa 11954  
Ile Pro Phe Phe Ala Ala Phe Cys Leu Pro Val Phe Ala His Pro Glu  
2035 2040 2045  
  
acg ctg gtg aaa gta aaa gat gct gaa gat cag ttg ggt gca cga gtg 12002  
Thr Leu Val Lys Val Lys Asp Ala Glu Asp Gln Leu Gly Ala Arg Val  
2050 2055 2060  
  
ggt tac atc gaa ctg gat ctc aac agc ggt aag atc ctt gag agt ttt 12050  
Gly Tyr Ile Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe  
2065 2070 2075  
  
cgc ccc gaa gaa cgt ttt cca atg atg agc act ttt aaa gtt ctg cta 12098  
Arg Pro Glu Glu Arg Phe Pro Met Met Ser Thr Phe Lys Val Leu Leu  
2080 2085 2090 2095  
  
tgt ggc gcg gta tta tcc cgt att gac gcc ggg caa gag caa ctc ggt 12146  
Cys Gly Ala Val Leu Ser Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly  
2100 2105 2110

cgc cgc ata cac tat tct cag aat gac ttg gtt gag tac tca cca gtc 12194  
 Arg Arg Ile His Tyr Ser Gln Asn Asp Leu Val Glu Tyr Ser Pro Val  
 2115 2120 2125

aca gaa aag cat ctt acg gat ggc atg aca gta aga gaa tta tgc agt 12242  
 Thr Glu Lys His Leu Thr Asp Gly Met Thr Val Arg Glu Leu Cys Ser  
 2130 2135 2140

gct gcc ata acc atg agt gat aac act gcg gcc aac tta ctt ctg aca 12290  
 Ala Ala Ile Thr Met Ser Asp Asn Thr Ala Ala Asn Leu Leu Thr  
 2145 2150 2155

acg atc gga gga ccg aag gag cta acc gct ttt ttg cac aac atg ggg 12338  
 Thr Ile Gly Gly Pro Lys Glu Leu Thr Ala Phe Leu His Asn Met Gly  
 2160 2165 2170 2175

gat cat gta act cgc ctt gat cgt tgg gaa ccg gag ctg aat gaa gcc 12386  
 Asp His Val Thr Arg Leu Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala  
 2180 2185 2190

ata cca aac gac gag cgt gac acc acg atg cct gta gca atg gca aca 12434  
 Ile Pro Asn Asp Glu Arg Asp Thr Thr Met Pro Val Ala Met Ala Thr  
 2195 2200 2205

acg ttg cgc aaa cta tta act ggc gaa cta ctt act cta gct tcc ccg 12482  
 Thr Leu Arg Lys Leu Leu Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg  
 2210 2215 2220

caa caa tta ata gac tgg atg gag gcg gat aaa gtt gca gga cca ctt 12530  
 Gln Gln Leu Ile Asp Trp Met Glu Ala Asp Lys Val Ala Gly Pro Leu  
 2225 2230 2235

ctg cgc tcg gcc ctt ccg gct ggc tgg ttt att gct gat aaa tct gga 12578  
 Leu Arg Ser Ala Leu Pro Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly  
 2240 2245 2250 2255

gcc ggt gag cgt ggg tct cgc ggt atc att gca gca ctg ggg cca gat 12626  
 Ala Gly Glu Arg Gly Ser Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp  
 2260 2265 2270

ggt aag ccc tcc cgt atc gta gtt atc tac acg acg ggg agt cag gca 12674  
 Gly Lys Pro Ser Arg Ile Val Val Ile Tyr Thr Gly Ser Gln Ala  
 2275 2280 2285

act atg gat gaa cga aat aga cag atc gct gag ata ggt gcc tca ctg 12722  
 Thr Met Asp Glu Arg Asn Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu  
 2290 2295 2300

att aag cat tgg taactgtcag accaagttta ctcatatata cttagatttg 12774  
 Ile Lys His Trp  
 2305

attnaaaaact tcattttaa tttnnnnn gataatctca 12834

tgacccaaaat cccttaacgt gagtttcgt tccactgagc gtcagacccc gtagaaaaaga 12894

tcaaaggatc ttctttagat ccttttttc tgccgtaat ctgctgcttg caaaacaaaaaa 12954

aaccaccgct accagcggtg gtttgttgc cggatcaaga gctaccaact cttttccga 13014

aggtaactgg cttcagcaga gcgagatac caaatactgt cttcttagtg tagccgtagt 13074

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 cgcttccga agggagaaaag gcggacaggt atccggtaag cggcagggtc ggaacaggag 13374  
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 aaaacgcac caacgcggcc ttttacggc tcctggcctt ttgctggcct tttgctcaca 13554  
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 aagagcgc ccc aatacgcaaa ccgcctctcc ccgcgcgttg gccgattcat taatgcagct 13734  
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 gaattgtgag cggataacaa tttcacacag gaaacagcta tgaccatgat tacgcc 13910

&lt;210&gt; 9

&lt;211&gt; 2307

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;400&gt; 9

Met	Asn	Gly	Gly	His	Ile	Gln	Leu	Ile	Ile	Gly	Pro	Met	Phe	Ser	Gly
1					5				10						15

Lys	Ser	Thr	Glu	Leu	Ile	Arg	Arg	Val	Arg	Arg	Tyr	Gln	Ile	Ala	Gln
					20			25					30		

Tyr	Lys	Cys	Val	Thr	Ile	Lys	Tyr	Ser	Asn	Asp	Asn	Arg	Tyr	Gly	Thr
					35			40				45			

Gly	Leu	Trp	Thr	His	Asp	Lys	Asn	Asn	Phe	Glu	Ala	Leu	Glu	Ala	Thr
					50		55				60				

Lys	Leu	Cys	Asp	Val	Leu	Glu	Ser	Ile	Thr	Asp	Phe	Ser	Val	Ile	Gly
					65		70		75				80		

Ile	Asp	Glu	Gly	Gln	Phe	Phe	Pro	Asp	Ile	Val	Glu	Met	Gly	Ile	Pro
					85			90				95			

Gln	Phe	Met	Ala	Arg	Val	Cys	Ala	Cys	Leu	Trp	Met	Met	Leu	Leu	Ile
					100			105				110			

Ala	Gln	Ala	Glu	Ala	Ala	Leu	Glu	Asn	Leu	Val	Val	Leu	Asn	Ala	Ala
					115			120				125			

Ser Val Ala Gly Ala His Gly Ile Leu Ser Phe Leu Val Phe Phe Cys  
 130 135 140  
 Ala Ala Trp Tyr Ile Lys Gly Arg Leu Val Pro Gly Ala Ala Tyr Ala  
 145 150 155 160  
 Leu Tyr Gly Val Trp Pro Leu Leu Leu Leu Ala Leu Pro Pro  
 165 170 175  
 Arg Ala Tyr Ala Met Asp Arg Glu Met Ala Ala Ser Cys Gly Gly Ala  
 180 185 190  
 Val Phe Val Gly Leu Val Leu Leu Thr Leu Ser Pro Tyr Tyr Lys Val  
 195 200 205  
 Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr Phe Thr Thr Arg Ala  
 210 215 220  
 Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn Ala Arg Gly Gly  
 225 230 235 240  
 Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His Pro Glu Leu Ile  
 245 250 255  
 Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly Pro Leu Met Val  
 260 265 270  
 Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val Arg Ala Gln Gly  
 275 280 285  
 Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala Gly Gly His Tyr  
 290 295 300  
 Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr Gly Thr Tyr Ile  
 305 310 315 320  
 Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His Ala Gly Leu Arg  
 325 330 335  
 Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser Asp Met Glu Thr  
 340 345 350  
 Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala Cys Gly Asp Ile Ile  
 355 360 365  
 Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu Ile Leu Leu Gly  
 370 375 380  
 Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu Leu Ala Pro Ile  
 385 390 395 400  
 Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly Cys Ile Ile Thr  
 405 410 415  
 Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly Glu Val Gln Val  
 420 425 430  
 Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys Val Asn Gly Val  
 435 440 445

Cys Trp Thr Val Tyr His Gly Ala Gly Ser Lys Thr Leu Ala Gly Pro  
   450                          455                          460  
  
 Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp Gln Asp Leu Val  
   465                          470                          475                          480  
  
 Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr Pro Cys Thr Cys  
   485                          490                          495  
  
 Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala Asp Val Ile Pro  
   500                          505                          510  
  
 Val Arg Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu Ser Pro Arg Pro  
   515                          520                          525  
  
 Val Ser Tyr Leu Lys Gly Ser Ala Gly Gly Pro Leu Leu Cys Pro Ser  
   530                          535                          540  
  
 Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys Thr Arg Gly Val  
   545                          550                          555                          560  
  
 Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met Glu Thr Thr Met  
   565                          570                          575  
  
 Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro Ala Val Pro Gln  
   580                          585                          590  
  
 Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly Ser Gly Lys Ser  
   595                          600                          605  
  
 Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr Lys Val Leu Val  
   610                          615                          620  
  
 Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly Ala Tyr Met Ser  
   625                          630                          635                          640  
  
 Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly Val Arg Thr Ile  
   645                          650                          655  
  
 Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly Lys Phe Leu Ala  
   660                          665                          670  
  
 Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile Ile Cys Asp Glu  
   675                          680                          685  
  
 Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile Gly Thr Val Leu  
   690                          695                          700  
  
 Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val Leu Ala Thr Ala  
   705                          710                          715                          720  
  
 Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn Ile Glu Glu Val  
   725                          730                          735  
  
 Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly Lys Ala Ile Pro  
   740                          745                          750  
  
 Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe Cys His Ser Lys  
   755                          760                          765

Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly Leu Gly Ile Asn  
 770 775 780  
 Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Val Ile Pro Thr Ile  
 785 790 795 800  
 Gly Asp Val Val Val Ala Thr Asp Ala Leu Met Thr Gly Tyr Thr  
 805 810 815  
 Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys Val Thr Gln Thr  
 820 825 830  
 Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu Thr Thr Thr Val  
 835 840 845  
 Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly Arg Thr Gly Arg  
 850 855 860  
 Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly Glu Arg Pro Ser  
 865 870 875 880  
 Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr Asp Ala Gly Cys  
 885 890 895  
 Ala Trp Tyr Glu Leu Thr Pro Ala Glu Thr Ser Val Arg Leu Arg Ala  
 900 905 910  
 Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp His Leu Glu Phe  
 915 920 925  
 Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp Ala His Phe Leu  
 930 935 940  
 Ser Gln Thr Lys Gln Ala Gly Asp Asn Phe Pro Tyr Leu Val Ala Tyr  
 945 950 955 960  
 Gln Ala Thr Val Cys Ala Arg Ala Gln Ala Pro Pro Pro Ser Trp Asp  
 965 970 975  
 Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr Leu His Gly Pro  
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Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala Ala Ala Pro Val  
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Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp Ala Lys His Met  
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Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe Thr Ala Ser Ile  
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Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe Asn Ile Leu Gly  
 1170 1175 1180

Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser Ala Ala Ser Ala Phe  
 185 1190 1195 1200

Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly Ser Ile Gly Leu Gly  
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Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala Gly Val Ala Gly  
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Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met Pro Ser Thr Glu  
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Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly Ala Leu Glu Leu  
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Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu Ser Leu Gly Ile  
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 Thr Val Cys Ile Ser Ser Tyr Asp His Asp Asn Gln Arg Glu Leu Lys  
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 Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys Met Lys  
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 Cys Tyr Ile Asp Ser Met Ser Ile Gln His Phe Arg Val Ala Leu Ile  
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 Lys Pro Ser Arg Ile Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr  
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Gly Leu Trp Thr His Asp Lys Asn Asn Phe Glu Ala Leu Glu Ala Thr  
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Leu Asn Ala Ala Ser Val Ala Gly Ala His Gly Ile Leu Ser Phe Leu  
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Val Phe Phe Cys Ala Ala Trp Tyr Ile Lys Gly Arg Leu Val Pro Gly  
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Ala Ala Tyr Ala Leu Tyr Gly Val Trp Pro Leu Leu Leu Leu Leu  
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Ala Leu Pro Pro Arg Ala Tyr Ala Met Asp Arg Glu Met Ala Ala Ser  
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Cys Gly Gly Ala Val Phe Val Gly Leu Val Leu Leu Thr Leu Ser Pro  
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Tyr Tyr Lys Val Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr Phe  
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Thr Thr Arg Ala Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn  
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Ala Arg Gly Gly Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His  
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Pro Glu Leu Ile Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly  
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Pro Leu Met Val Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val  
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Arg Ala Gln Gly Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala  
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Gly Gly His Tyr Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr  
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Gly Thr Tyr Ile Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His  
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Ala Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser  
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Gly Asp Ile Ile Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu  
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Ile Leu Leu Gly Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu  
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Leu Ala Pro Ile Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly  
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Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly  
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Ser Pro Arg Pro Val Ser Tyr Leu Lys Gly Ser Ala Gly Gly Pro Leu  
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Leu Cys Pro Ser Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys  
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Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr  
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Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly  
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Ala Tyr Met Ser Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly  
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Val Arg Thr Ile Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly  
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Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile  
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Ile Cys Asp Glu Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile  
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Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val  
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Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn  
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Ile Glu Glu Val Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly  
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Lys Ala Ile Pro Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe  
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Cys His Ser Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly  
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Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu  
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His Leu Glu Phe Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp  
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 Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr  
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Val Ser Thr Val Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp  
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 Glu Leu Gly Ile Arg His Val Asp Thr Val Cys Ile Ser Ser Tyr Asp  
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 His Asp Asn Gln Arg Glu Leu Lys Val Leu Lys Arg Ala Glu Gly Asp  
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 Ala Val Ala Ile Arg Glu Met Tyr Pro Lys Ala His Phe Val Thr Ile  
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<223> E. coli gpt; for selection of recombinants

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aga gcc gag gcg cac tta cat gtg tgg atc ccc ccc ctc aac gct cg <sup>g</sup> Arg Ala Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn Ala Arg 225 230 235	1862
gga ggc cgc gat gcc atc atc ctc atc atg tgc gca gtc cat cca gag Gly Gly Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His Pro Glu 240 245 250	1910
cta atc ttt gac atc acc aaa ctt cta att gcc ata ctc ggt ccg ctc Leu Ile Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly Pro Leu 255 260 265 270	1958
atg gtg ctc caa gct ggc ata acc aga gtg ccg tac ttc gtg cg <sup>c</sup> gct Met Val Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val Arg Ala 275 280 285	2006
caa ggg ctc att cat gca tgc atg tta gtg ccg aag gtc gct ggg ggt Gln Gly Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala Gly Gly 290 295 300	2054
cat tat gtc caa atg gcc ttc atg aag ctg ggc gcg ctg aca ggc acg His Tyr Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr Gly Thr 305 310 315	2102
tac att tac aac cat ctt acc ccg cta ccg gat tgg gcc cac gcg ggc Tyr Ile Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His Ala Gly 320 325 330	2150
cta cga gac ctt gc <sup>g</sup> gca gtg gag ccc gtc gtc ttc tcc gac atg Leu Arg Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser Asp Met 335 340 345 350	2198
gag acc aag atc atc acc tgg gga gca gac acc gcg gcg gct ggg gac Glu Thr Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala Ala Gly Asp 355 360 365	2246
atc atc ttg ggt ctg ccc gtc tcc gcc cga agg gga aag gag ata ctc Ile Ile Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu Ile Leu 370 375 380	2294
ctg ggc ccg gcc gat agt ctt gaa ggg ccg ggg tgg cga ctc ctc gcg Leu Gly Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu Leu Ala 385 390 395	2342
ccc atc acg gcc tac tcc caa cag acg ccg ggc cta ctt ggt tgc atc Pro Ile Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly Cys Ile 400 405 410	2390

atc act agc ctt aca ggc cg <sup>g</sup> gac aag aac cag gtc gag gga gag gtt Ile Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly Glu Val 415 420 425 430	2438
cag gtg gtt tcc acc gca aca caa tcc ttc ctg gcg acc tgc gtc aac Gln Val Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys Val Asn 435 440 445	2486
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ggc cca aag ggg cca atc acc cag atg tac act aat gtg gac cag gac Gly Pro Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp Gln Asp 465 470 475	2582
ctc gtc ggc tgg cag gcg ccc ccc ggg gcg cgt tcc ttg aca cca tgc Leu Val Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr Pro Cys 480 485 490	2630
acc tgt ggc agc tca gac ctt tac ttg gtc acg aga cat gct gac gtc Thr Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala Asp Val 495 500 505 510	2678
att ccg gtg cgc cgg cgg ggc gac agt agg ggg agc ctg ctc tcc ccc Ile Pro Val Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu Ser Pro 515 520 525	2726
agg cct gtc tcc tac ttg aag ggc tct gcg ggt ggt cca ctg ctc tgc Arg Pro Val Ser Tyr Leu Lys Gly Ser Ala Gly Pro Leu Leu Cys 530 535 540	2774
cct tcg ggg cac gct gtg ggc atc ttc cgg gct gcc gta tgc acc cgg Pro Ser Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys Thr Arg 545 550 555	2822
ggg gtt gcg aag gcg gtg gac ttt gtg ccc gta gag tcc atg gaa act Gly Val Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met Glu Thr 560 565 570	2870
act atg cgg tct ccg gtc ttc acg gac aac tca tcc ccc ccg gcc gta Thr Met Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro Ala Val 575 580 585 590	2918
ccg cag tca ttt caa gtg gcc cac cta cac gct ccc act ggc agc ggc Pro Gln Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly Ser Gly 595 600 605	2966
aag agt act aaa gtg ccg gct gca tat gca gcc caa ggg tac aag gtg Lys Ser Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr Lys Val 610 615 620	3014
ctc gtc ctc aat ccg tcc gtt gcc gct acc tta ggg ttt ggg gcg tat Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly Ala Tyr 625 630 635	3062
atg tct aag gca cac ggt att gac ccc aac atc aga act ggg gta agg Met Ser Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly Val Arg 640 645 650	3110

acc att acc aca ggc gcc ccc gtc aca tac tct acc tat ggc aag ttt		3158
Thr Ile Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly Lys Phe		
655 660 665 670		
ctt gcc gat ggt ggt tgc tct ggg ggc gct tat gac atc ata ata tgt		3206
Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile Cys		
675 680 685		
gat gag tgc cat tca act gac tcg act aca atc ttg ggc atc ggc aca		3254
Asp Glu Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile Gly Thr		
690 695 700		
gtc ctg gac caa gcg gag acg gct gga gcg cgg ctt gtc gtg ctc gcc		3302
Val Leu Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val Leu Ala		
705 710 715		
acc gct acg cct ccg gga tcg gtc acc gtg cca cac cca aac atc gag		3350
Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn Ile Glu		
720 725 730		
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Glu Val Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly Lys Ala		
735 740 745 750		
atc ccc att gaa gcc atc agg ggg gga agg cat ctc att ttc tgt cat		3446
Ile Pro Ile Glu Ala Ile Arg Gly Arg His Leu Ile Phe Cys His		
755 760 765		
tcc aag aag aag tgc gac gag ctc gcc gca aag ctg tca ggc ctc gga		3494
Ser Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly Leu Gly		
770 775 780		
atc aac gct gtg gcg tat tac cgg ggg ctc gat gtg tcc gtc ata cca		3542
Ile Asn Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Val Ile Pro		
785 790 795		
act atc gga gac gtc gtt gtc gtg gca aca gac gct ctg atg acg ggc		3590
Thr Ile Gly Asp Val Val Val Ala Thr Asp Ala Leu Met Thr Gly		
800 805 810		
tat acg ggc gac ttt gac tca gtg atc gac tgt aac aca tgt gtc acc		3638
Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys Val Thr		
815 820 825 830		
cag aca gtc gac ttc agc ttg gat ccc acc ttc acc att gag acg acg		3686
Gln Thr Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu Thr Thr		
835 840 845		
acc gtg cct caa gac gca gtg tcg cgc tcg cag cgg cgg ggt agg act		3734
Thr Val Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly Arg Thr		
850 855 860		
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Gly Arg Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly Glu Arg		
865 870 875		
ccc tcg ggc atg ttc gat tcc tcg gtc ctg tgt gag tgc tat gac gcg		3830
Pro Ser Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr Asp Ala		
880 885 890		

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cg <sup>g</sup> gcc tac ctg aac aca cca ggg ttg ccc gtt tgc cag gac cac ctg Arg Ala Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp His Leu 915 920 925	3926
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tgg gat caa atg tgg aag tgt ctc ata cgg ctg aaa cct acg ctg cac Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr Leu His 975 980 985 990	4118
ggg cca aca ccc ttg ctg tac agg ctg gga gcc gtc caa aat gag gtc Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ala Val Gln Asn Glu Val 995 1000 1005	4166
acc ctc acc cac ccc ata acc aaa tac atc atg gca tgc atg tcg gct Thr Leu Thr His Pro Ile Thr Lys Tyr Ile Met Ala Cys Met Ser Ala 1010 1015 1020	4214
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gca gct ctg gcc gcg tat tgc ctg aca aca ggc agt gtg gtc att gtg Ala Ala Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val Val Ile Val 1040 1045 1050	4310
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cct tac atc gag cag gga atg cag ctc gcc gag caa ttc aag cag aaa Pro Tyr Ile Glu Gln Gly Met Gln Leu Ala Glu Gln Phe Lys Gln Lys 1090 1095 1100	4454
g <sup>cg</sup> ctc ggg tta ctg caa aca gcc acc aaa caa gcg gag gct gct gct Ala Leu Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala Ala Ala 1105 1110 1115	4502
ccc gtg gtg gag tcc aag tgg cga gcc ctt gag aca ttc tgg gcg aag Pro Val Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp Ala Lys 1120 1125 1130	4550

cac atg tgg aat ttc atc agc ggg ata cag tac tta gca ggc tta tcc His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly Leu Ser 1135 1140 1145 1150	4598
act ctg cct ggg aac ccc gca ata gca tca ttg atg gca ttc aca gcc Thr Leu Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe Thr Ala 1155 1160 1165	4646
tct atc acc agc ccg ctc acc acc caa agt acc ctc ctg ttt aac atc Ser Ile Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe Asn Ile 1170 1175 1180	4694
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ctt ggg aag gtg ctt gtg gac att ctg gcg ggt tat gga gca gga gtg Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala Gly Val 1215 1220 1225 1230	4838
gcc ggc gcg ctc gtg gcc ttt aag gtc atg agc ggc gag atg ccc tcc Ala Gly Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met Pro Ser 1235 1240 1245	4886
acc gag gac ctg gtc aat cta ctt cct gcc atc ctc gag gaa gct agt Thr Glu Asp Leu Val Asn Leu Leu Pro Ala Ile Leu Glu Glu Ala Ser 1250 1255 1260	4934
gag gat gtc gtc tgc tca atg tcc tac aca tgg aca ggc gcc ttg Glu Asp Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly Ala Leu 1265 1270 1275	4982
gag ctg ctg ctg ctg ctg ggc ctg agg cta cag ctc tcc ctg Glu Leu Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu Ser Leu 1280 1285 1290	5030
ggc atc atc cca gtt gag gag aac ccg gac ttc tgg aac cgc gag Gly Ile Ile Pro Val Glu Glu Asn Pro Asp Phe Trp Asn Arg Glu 1295 1300 1305 1310	5078
gca gcc gag gcc ggt gcc gcc aag aag ctg cag cct gca cag aca Ala Ala Glu Ala Ala Lys Lys Leu Gln Pro Ala Gln Thr 1315 1320 1325	5126
gcc gcc aag aac ctc atc atc ttc ctg ggc gat ggg atg ggg gtg tct Ala Ala Lys Asn Leu Ile Ile Phe Leu Gly Asp Gly Met Gly Val Ser 1330 1335 1340	5174
acg gtg aca gct gcc agg atc cta aaa ggg cag aag aag gac aaa ctg Thr Val Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp Lys Leu 1345 1350 1355	5222
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gga gac atg aaa tac gag atc cac cga gac tcc aca ctg gac ccc tcc Gly Asp Met Lys Tyr Glu Ile His Arg Asp Ser Thr Leu Asp Pro Ser 1570 1575 1580	5894
ctg atg gag atg aca gag gct gcc ctg cgc ctg ctg agc agg aac ccc Leu Met Glu Met Thr Glu Ala Ala Leu Arg Leu Leu Ser Arg Asn Pro 1585 1590 1595	5942
cgc ggc ttc ttc ctc ttc gtg gag ggt ggt cgc atc gac cat ggt cat Arg Gly Phe Phe Leu Phe Val Glu Gly Gly Arg Ile Asp His Gly His 1600 1605 1610	5990



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 1785 1790 1795  
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 Leu Gln Ile His Ala Arg Lys Leu Ala Ser Arg Leu Met Pro Ser Glu  
 1800 1805 1810

caa tgg aaa ggc att att gcc gta agc cgt ggc ggt ctg gta ccg ggt		8707
Gln Trp Lys Gly Ile Ile Ala Val Ser Arg Gly Gly Leu Val Pro Gly		
1815	1820	1825
gcg tta ctg gcg cgt gaa ctg ggt att cgt cat gtc gat acc gtt tgt		8755
Ala Leu Leu Ala Arg Glu Leu Gly Ile Arg His Val Asp Thr Val Cys		
1830	1835	1840
att tcc agc tac gat cac gac aac cag cgc gag ctt aaa gtg ctg aaa		8803
Ile Ser Ser Tyr Asp His Asp Asn Gln Arg Glu Leu Lys Val Leu Lys		
1845	1850	1855
cgc gca gaa ggc gat ggc gaa ggc ttc atc gtt att gat gac ctg gtg		8851
Arg Ala Glu Gly Asp Gly Glu Gly Phe Ile Val Ile Asp Asp Leu Val		
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Asp Thr Gly Thr Ala Val Ala Ile Arg Glu Met Tyr Pro Lys Ala		
1880	1885	1890
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His Phe Val Thr Ile Phe Ala Lys Pro Ala Gly Arg Pro Leu Val Asp		
1895	1900	1905
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Asp Tyr Val Val Asp Ile Pro Gln Asp Thr Trp Ile Glu Gln Pro Trp		
1910	1915	1920
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Asp Met Gly Val Val Phe Val Pro Pro Ile Ser Gly Arg		
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Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val  
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Ala Ala Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu  
1955 1960 1965  
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Asn Leu Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys  
1970 1975 1980  
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Met Lys Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Gly Glu Glu  
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Thr Glu Ile Glu Ile Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys  
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acaqttgc当地 11315

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taatattgaa aaaggaagag t atg agt att caa cat ttc cgt gtc gcc ctt 11906  
Met Ser Ile His Phe Arg Val Ala Leu  
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Ile Pro Phe Phe Ala Ala Phe Cys Leu Pro Val Phe Ala His Pro Glu  
2035 2040 2045  
acg ctg gtg aaa gta aaa gat gct gaa gat cag ttg ggt gca cga gtg 12002  
Thr Leu Val Lys Val Lys Asp Ala Glu Asp Gln Leu Gly Ala Arg Val  
2050 2055 2060  
ggt tac atc gaa ctg gat ctc aac agc ggt aag atc ctt gag agt ttt 12050  
Gly Tyr Ile Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe  
2065 2070 2075  
cgc ccc gaa gaa cgt ttt cca atg atg agc act ttt aaa gtt ctg cta 12098  
Arg Pro Glu Glu Arg Phe Pro Met Met Ser Thr Phe Lys Val Leu Leu  
2080 2085 2090 2095  
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Cys Gly Ala Val Leu Ser Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly  
2100 2105 2110  
cgc cgc ata cac tat tct cag aat gac ttg gtt gag tac tca cca gtc 12194  
Arg Arg Ile His Tyr Ser Gln Asn Asp Leu Val Glu Tyr Ser Pro Val  
2115 2120 2125  
aca gaa aag cat ctt acg gat ggc atg aca gta aga gaa tta tgc agt 12242  
Thr Glu Lys His Leu Thr Asp Gly Met Thr Val Arg Glu Leu Cys Ser  
2130 2135 2140  
gct gcc ata acc atg agt gat aac act gcg gcc aac tta ctt ctg aca 12290  
Ala Ala Ile Thr Met Ser Asp Asn Thr Ala Ala Asn Leu Leu Thr  
2145 2150 2155  
acg atc gga gga ccg aag gag cta acc gct ttt ttg cac aac atg ggg 12338  
Thr Ile Gly Gly Pro Lys Glu Leu Thr Ala Phe Leu His Asn Met Gly  
2160 2165 2170 2175  
gat cat gta act cgc ctt gat cgt tgg gaa ccg gag ctg aat gaa gcc 12386  
Asp His Val Thr Arg Leu Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala  
2180 2185 2190

ata cca aac gac gag cgt gac acc acg atg cct gta gca atg gca aca Ile Pro Asn Asp Glu Arg Asp Thr Thr Met Pro Val Ala Met Ala Thr 2195	2200	2205	12434	
acg ttg cgc aaa cta tta act ggc gaa cta ctt act cta gct tcc cgg Thr Leu Arg Lys Leu Leu Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg 2210	2215	2220	12482	
caa caa tta ata gac tgg atg gag gcg gat aaa gtt gca gga cca ctt Gln Gln Leu Ile Asp Trp Met Glu Ala Asp Lys Val Ala Gly Pro Leu 2225	2230	2235	12530	
ctg cgc tcg gcc ctt ccg gct ggc tgg ttt att gct gat aaa tct gga Leu Arg Ser Ala Leu Pro Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly 2240	2245	2250	2255	12578
gcc ggt gag cgt ggg tct cgc ggt atc att gca gca ctg ggg cca gat Ala Gly Glu Arg Gly Ser Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp 2260	2265	2270	12626	
ggt aag ccc tcc cgt atc gta gtt atc tac acg acg ggg agt cag gca Gly Lys Pro Ser Arg Ile Val Val Ile Tyr Thr Thr Gly Ser Gln Ala 2275	2280	2285	12674	
act atg gat gaa cga aat aga cag atc gct gag ata ggt gcc tca ctg Thr Met Asp Glu Arg Asn Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu 2290	2295	2300	12722	
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			13314	
			13374	
			13434	
			13494	
			13554	
			13614	

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 aagagcgccc aatacgcaaa ccgcctctcc ccgcgcgttg gccgattcat taatgcagct 13734  
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 <211> 2307  
 <212> PRT  
 <213> Artificial Sequence

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 Tyr Lys Cys Val Thr Ile Lys Tyr Ser Asn Asp Asn Arg Tyr Gly Thr  
 35 40 45  
 Gly Leu Trp Thr His Asp Lys Asn Asn Phe Glu Ala Leu Glu Ala Thr  
 50 55 60  
 Lys Leu Cys Asp Val Leu Glu Ser Ile Thr Asp Phe Ser Val Ile Gly  
 65 70 75 80  
 Ile Asp Glu Gly Gln Phe Phe Pro Asp Ile Val Glu Met Gly Ile Pro  
 85 90 95  
 Gln Phe Met Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu Leu Ile  
 100 105 110  
 Ala Gln Ala Glu Ala Ala Leu Glu Asn Leu Val Val Leu Asn Ala Ala  
 115 120 125  
 Ser Val Ala Gly Ala His Gly Ile Leu Ser Phe Leu Val Phe Phe Cys  
 130 135 140  
 Ala Ala Trp Tyr Ile Lys Gly Arg Leu Val Pro Gly Ala Ala Tyr Ala  
 145 150 155 160  
 Leu Tyr Gly Val Trp Pro Leu Leu Leu Leu Leu Ala Leu Pro Pro  
 165 170 175  
 Arg Ala Tyr Ala Met Asp Arg Glu Met Ala Ala Ser Cys Gly Gly Ala  
 180 185 190  
 Val Phe Val Gly Leu Val Leu Leu Thr Leu Ser Pro Tyr Tyr Lys Val  
 195 200 205  
 Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr Phe Thr Thr Arg Ala  
 210 215 220  
 Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn Ala Arg Gly Gly  
 225 230 235 240

Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His Pro Glu Leu Ile  
 245 250 255  
 Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly Pro Leu Met Val  
 260 265 270  
 Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val Arg Ala Gln Gly  
 275 280 285  
 Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala Gly Gly His Tyr  
 290 295 300  
 Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr Gly Thr Tyr Ile  
 305 310 315 320  
 Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His Ala Gly Leu Arg  
 325 330 335  
 Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser Asp Met Glu Thr  
 340 345 350  
 Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala Ala Gly Asp Ile Ile  
 355 360 365  
 Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu Ile Leu Leu Gly  
 370 375 380  
 Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu Leu Ala Pro Ile  
 385 390 395 400  
 Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly Cys Ile Ile Thr  
 405 410 415  
 Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly Glu Val Gln Val  
 420 425 430  
 Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys Val Asn Gly Val  
 435 440 445  
 Cys Trp Thr Val Tyr His Gly Ala Gly Ser Lys Thr Leu Ala Gly Pro  
 450 455 460  
 Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp Gln Asp Leu Val  
 465 470 475 480  
 Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr Pro Cys Thr Cys  
 485 490 495  
 Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala Asp Val Ile Pro  
 500 505 510  
 Val Arg Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu Ser Pro Arg Pro  
 515 520 525  
 Val Ser Tyr Leu Lys Gly Ser Ala Gly Gly Pro Leu Leu Cys Pro Ser  
 530 535 540  
 Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys Thr Arg Gly Val  
 545 550 555 560

Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met Glu Thr Thr Met  
 565 570 575  
 Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro Ala Val Pro Gln  
 580 585 590  
 Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly Ser Gly Lys Ser  
 595 600 605  
 Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr Lys Val Leu Val  
 610 615 620  
 Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly Ala Tyr Met Ser  
 625 630 635 640  
 Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly Val Arg Thr Ile  
 645 650 655  
 Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly Lys Phe Leu Ala  
 660 665 670  
 Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile Ile Cys Asp Glu  
 675 680 685  
 Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile Gly Thr Val Leu  
 690 695 700  
 Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val Leu Ala Thr Ala  
 705 710 715 720  
 Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn Ile Glu Glu Val  
 725 730 735  
 Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly Lys Ala Ile Pro  
 740 745 750  
 Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe Cys His Ser Lys  
 755 760 765  
 Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly Leu Gly Ile Asn  
 770 775 780  
 Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Val Ile Pro Thr Ile  
 785 790 795 800  
 Gly Asp Val Val Val Ala Thr Asp Ala Leu Met Thr Gly Tyr Thr  
 805 810 815  
 Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys Val Thr Gln Thr  
 820 825 830  
 Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu Thr Thr Val  
 835 840 845  
 Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly Arg Thr Gly Arg  
 850 855 860  
 Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly Glu Arg Pro Ser  
 865 870 875 880

Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr Asp Ala Gly Cys  
 885 890 895  
 Ala Trp Tyr Glu Leu Thr Pro Ala Glu Thr Ser Val Arg Leu Arg Ala  
 900 905 910  
 Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp His Leu Glu Phe  
 915 920 925  
 Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp Ala His Phe Leu  
 930 935 940  
 Ser Gln Thr Lys Gln Ala Gly Asp Asn Phe Pro Tyr Leu Val Ala Tyr  
 945 950 955 960  
 Gln Ala Thr Val Cys Ala Arg Ala Gln Ala Pro Pro Pro Ser Trp Asp  
 965 970 975  
 Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr Leu His Gly Pro  
 980 985 990  
 Thr Pro Leu Leu Tyr Arg Leu Gly Ala Val Gln Asn Glu Val Thr Leu  
 995 1000 1005  
 Thr His Pro Ile Thr Lys Tyr Ile Met Ala Cys Met Ser Ala Asp Leu  
 1010 1015 1020  
 Glu Val Val Thr Ser Thr Trp Val Leu Val Gly Gly Val Leu Ala Ala  
 1025 1030 1035 1040  
 Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val Val Ile Val Gly Arg  
 1045 1050 1055  
 Ile Ile Leu Ser Gly Arg Pro Ala Ile Val Pro Asp Arg Glu Leu Leu  
 1060 1065 1070  
 Tyr Gln Glu Phe Asp Glu Met Glu Glu Cys Ala Ser His Leu Pro Tyr  
 1075 1080 1085  
 Ile Glu Gln Gly Met Gln Leu Ala Glu Gln Phe Lys Gln Lys Ala Leu  
 1090 1095 1100  
 Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala Ala Ala Pro Val  
 1095 1110 1115 1120  
 Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp Ala Lys His Met  
 1125 1130 1135  
 Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly Leu Ser Thr Leu  
 1140 1145 1150  
 Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe Thr Ala Ser Ile  
 1155 1160 1165  
 Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe Asn Ile Leu Gly  
 1170 1175 1180  
 Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser Ala Ala Ser Ala Phe  
 1185 1190 1195 1200

Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly Ser Ile Gly Leu Gly  
 1205 1210 1215  
 Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala Gly Val Ala Gly  
 1220 1225 1230  
 Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met Pro Ser Thr Glu  
 1235 1240 1245  
 Asp Leu Val Asn Leu Leu Pro Ala Ile Leu Glu Glu Ala Ser Glu Asp  
 1250 1255 1260  
 Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly Ala Leu Glu Leu  
 265 1270 1275 1280  
 Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu Ser Leu Gly Ile  
 1285 1290 1295  
 Ile Pro Val Glu Glu Asn Pro Asp Phe Trp Asn Arg Glu Ala Ala  
 1300 1305 1310  
 Glu Ala Leu Gly Ala Ala Lys Lys Leu Gln Pro Ala Gln Thr Ala Ala  
 1315 1320 1325  
 Lys Asn Leu Ile Ile Phe Leu Gly Asp Gly Met Gly Val Ser Thr Val  
 1330 1335 1340  
 Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp Lys Leu Gly Pro  
 345 1350 1355 1360  
 Glu Ile Pro Leu Ala Met Asp Arg Phe Pro Tyr Val Ala Leu Ser Lys  
 1365 1370 1375  
 Thr Tyr Asn Val Asp Lys His Val Pro Asp Ser Gly Ala Thr Ala Thr  
 1380 1385 1390  
 Ala Tyr Leu Cys Gly Val Lys Gly Asn Phe Gln Thr Ile Gly Leu Ser  
 1395 1400 1405  
 Ala Ala Ala Arg Phe Asn Gln Cys Asn Thr Thr Arg Gly Asn Glu Val  
 1410 1415 1420  
 Ile Ser Val Met Asn Arg Ala Lys Lys Ala Gly Lys Ser Val Gly Val  
 425 1430 1435 1440  
 Val Thr Thr Arg Val Gln His Ala Ser Pro Ala Gly Thr Tyr Ala  
 1445 1450 1455  
 His Thr Val Asn Arg Asn Trp Tyr Ser Asp Ala Asp Val Pro Ala Ser  
 1460 1465 1470  
 Ala Arg Gln Glu Gly Cys Gln Asp Ile Ala Thr Gln Leu Ile Ser Asn  
 1475 1480 1485  
 Met Asp Ile Asp Val Ile Leu Gly Gly Arg Lys Tyr Met Phe Pro  
 1490 1495 1500  
 Met Gly Thr Pro Asp Pro Glu Tyr Pro Asp Asp Tyr Ser Gln Gly Gly  
 505 1510 1515 1520

Thr Arg Leu Asp Gly Lys Asn Leu Val Gln Glu Trp Leu Ala Lys Arg  
 1525 1530 1535  
 Gln Gly Ala Arg Tyr Val Trp Asn Arg Thr Glu Leu Met Gln Ala Ser  
 1540 1545 1550  
 Leu Asp Pro Ser Val Thr His Leu Met Gly Leu Phe Glu Pro Gly Asp  
 1555 1560 1565  
 Met Lys Tyr Glu Ile His Arg Asp Ser Thr Leu Asp Pro Ser Leu Met  
 1570 1575 1580  
 Glu Met Thr Glu Ala Ala Leu Arg Leu Leu Ser Arg Asn Pro Arg Gly  
 585 1590 1595 1600  
 Phe Phe Leu Phe Val Glu Gly Arg Ile Asp His Gly His His Glu  
 1605 1610 1615  
 Ser Arg Ala Tyr Arg Ala Leu Thr Glu Thr Ile Met Phe Asp Asp Ala  
 1620 1625 1630  
 Ile Glu Arg Ala Gly Gln Leu Thr Ser Glu Glu Asp Thr Leu Ser Leu  
 1635 1640 1645  
 Val Thr Ala Asp His Ser His Val Phe Ser Phe Gly Gly Tyr Pro Leu  
 1650 1655 1660  
 Arg Gly Ser Cys Ile Phe Gly Leu Ala Pro Gly Lys Ala Arg Asp Arg  
 665 1670 1675 1680  
 Lys Ala Tyr Thr Val Leu Leu Tyr Gly Asn Gly Pro Gly Tyr Val Leu  
 1685 1690 1695  
 Lys Asp Gly Ala Arg Pro Asp Val Thr Glu Ser Glu Ser Gly Ser Pro  
 1700 1705 1710  
 Glu Tyr Arg Gln Gln Ser Ala Val Pro Leu Asp Glu Glu Thr His Ala  
 1715 1720 1725  
 Gly Glu Asp Val Ala Val Phe Ala Arg Gly Pro Gln Ala His Leu Val  
 1730 1735 1740  
 His Gly Val Gln Glu Gln Thr Phe Ile Ala His Val Met Ala Phe Ala  
 745 1750 1755 1760  
 Ala Cys Leu Glu Pro Tyr Thr Ala Cys Asp Leu Ala Pro Pro Ala Gly  
 1765 1770 1775  
 Thr Thr Asp Ala Ala His Pro Gly Met Ser Glu Lys Tyr Ile Val Thr  
 1780 785 1790  
 Trp Asp Met Leu Gln Ile His Ala Arg Lys Leu Ala Ser Arg Leu Met  
 1795 1800 1805  
 Pro Ser Glu Gln Trp Lys Gly Ile Ile Ala Val Ser Arg Gly Gly Leu  
 1810 1815 1820  
 Val Pro Gly Ala Leu Leu Ala Arg Glu Leu Gly Ile Arg His Val Asp  
 825 1830 1835 1840

Thr Val Cys Ile Ser Ser Tyr Asp His Asp Asn Gln Arg Glu Leu Lys  
 1845 1850 1855  
 Val Leu Lys Arg Ala Glu Gly Asp Gly Glu Gly Phe Ile Val Ile Asp  
 1860 1865 1870  
 Asp Leu Val Asp Thr Gly Gly Thr Ala Val Ala Ile Arg Glu Met Tyr  
 1875 1880 1885  
 Pro Lys Ala His Phe Val Thr Ile Phe Ala Lys Pro Ala Gly Arg Pro  
 1890 1895 1900  
 Leu Val Asp Asp Tyr Val Val Asp Ile Pro Gln Asp Thr Trp Ile Glu  
 905 1910 1915 1920  
 Gln Pro Trp Asp Met Gly Val Val Phe Val Pro Pro Ile Ser Gly Arg  
 1925 1930 1935  
 Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val Ala Ala  
 1940 1945 1950  
 Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu Asn Leu  
 1955 1960 1965  
 Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys Met Lys  
 1970 1975 1980  
 Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Gly Glu Glu Thr Glu  
 985 1990 1995 2000  
 Ile Glu Ile Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys Arg Lys  
 2005 2010 2015  
 Cys Tyr Ile Asp Ser Met Ser Ile Gln His Phe Arg Val Ala Leu Ile  
 2020 2025 2030  
 Pro Phe Phe Ala Ala Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr  
 2035 2040 2045  
 Leu Val Lys Val Lys Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly  
 2050 2055 2060  
 Tyr Ile Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg  
 065 2070 2075 208  
 Pro Glu Glu Arg Phe Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys  
 2085 2090 2095  
 Gly Ala Val Leu Ser Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly Arg  
 2100 2105 2110  
 Arg Ile His Tyr Ser Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr  
 2115 2120 2125  
 Glu Lys His Leu Thr Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala  
 2130 2135 2140  
 Ala Ile Thr Met Ser Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr  
 145 2150 2155 216

Ile Gly Gly Pro Lys Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp  
 2165 2170 2175

His Val Thr Arg Leu Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile  
 2180 2185 2190

Pro Asn Asp Glu Arg Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr  
 2195 2200 2205

Leu Arg Lys Leu Leu Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln  
 2210 2215 2220

Gln Leu Ile Asp Trp Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu  
 2225 2230 2235 224

Arg Ser Ala Leu Pro Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala  
 2245 2250 2255

Gly Glu Arg Gly Ser Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly  
 2260 2265 2270

Lys Pro Ser Arg Ile Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr  
 2275 2280 2285

Met Asp Glu Arg Asn Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile  
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Lys His Trp  
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<211> 92

<212> PRT

<213> Artificial Sequence

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Tyr Lys Cys Val Thr Ile Lys Tyr Ser Asn Asp Asn Arg Tyr Gly Thr  
 35 40 45

Gly Leu Trp Thr His Asp Lys Asn Asn Phe Glu Ala Leu Glu Ala Thr  
 50 55 60

Lys Leu Cys Asp Val Leu Glu Ser Ile Thr Asp Phe Ser Val Ile Gly  
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Ile Asp Glu Gly Gln Phe Phe Pro Asp Ile Val Glu  
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<210> 18

<211> 1692

<212> PRT

<213> Artificial Sequence

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 20 25 30

Leu Asn Ala Ala Ser Val Ala Gly Ala His Gly Ile Leu Ser Phe Leu  
 35 40 45

Val Phe Phe Cys Ala Ala Trp Tyr Ile Lys Gly Arg Leu Val Pro Gly  
 50 55 60

Ala Ala Tyr Ala Leu Tyr Gly Val Trp Pro Leu Leu Leu Leu Leu  
 65 70 75 80

Ala Leu Pro Pro Arg Ala Tyr Ala Met Asp Arg Glu Met Ala Ala Ser  
 85 90 95

Cys Gly Gly Ala Val Phe Val Gly Leu Val Leu Leu Thr Leu Ser Pro  
 100 105 110

Tyr Tyr Lys Val Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr Phe  
 115 120 125

Thr Thr Arg Ala Glu Ala His Leu His Val Trp Ile Pro Pro Leu Asn  
 130 135 140

Ala Arg Gly Gly Arg Asp Ala Ile Ile Leu Leu Met Cys Ala Val His  
 145 150 155 160

Pro Glu Leu Ile Phe Asp Ile Thr Lys Leu Leu Ile Ala Ile Leu Gly  
 165 170 175

Pro Leu Met Val Leu Gln Ala Gly Ile Thr Arg Val Pro Tyr Phe Val  
 180 185 190

Arg Ala Gln Gly Leu Ile His Ala Cys Met Leu Val Arg Lys Val Ala  
 195 200 205

Gly Gly His Tyr Val Gln Met Ala Phe Met Lys Leu Gly Ala Leu Thr  
 210 215 220

Gly Thr Tyr Ile Tyr Asn His Leu Thr Pro Leu Arg Asp Trp Ala His  
 225 230 235 240

Ala Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Val Val Phe Ser  
 245 250 255

Asp Met Glu Thr Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala Ala  
 260 265 270

Gly Asp Ile Ile Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Lys Glu  
 275 280 285

Ile Leu Leu Gly Pro Ala Asp Ser Leu Glu Gly Arg Gly Trp Arg Leu  
 290 295 300

Leu Ala Pro Ile Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly  
 305 310 315 320

Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly  
 325 330 335  
 Glu Val Gln Val Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys  
 340 345 350  
 Val Asn Gly Val Cys Trp Thr Val Tyr His Gly Ala Gly Ser Lys Thr  
 355 360 365  
 Leu Ala Gly Pro Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp  
 370 375 380  
 Gln Asp Leu Val Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr  
 385 390 395 400  
 Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala  
 405 410 415  
 Asp Val Ile Pro Val Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu  
 420 425 430  
 Ser Pro Arg Pro Val Ser Tyr Leu Lys Gly Ser Ala Gly Pro Leu  
 435 440 445  
 Leu Cys Pro Ser Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys  
 450 455 460  
 Thr Arg Gly Val Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met  
 465 470 475 480  
 Glu Thr Thr Met Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro  
 485 490 495  
 Ala Val Pro Gln Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly  
 500 505 510  
 Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr Ala Ala Gln Gly Tyr  
 515 520 525  
 Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly  
 530 535 540  
 Ala Tyr Met Ser Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly  
 545 550 555 560  
 Val Arg Thr Ile Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly  
 565 570 575  
 Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile  
 580 585 590  
 Ile Cys Asp Glu Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile  
 595 600 605  
 Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val  
 610 615 620  
 Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn  
 625 630 635 640

Ile Glu Glu Val Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly  
 645 650 655  
 Lys Ala Ile Pro Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe  
 660 665 670  
 Cys His Ser Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly  
 675 680 685  
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 Ile Pro Thr Ile Gly Asp Val Val Val Ala Thr Asp Ala Leu Met  
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 Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys  
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 Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Glu  
 740 745 750  
 Thr Thr Thr Val Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly  
 755 760 765  
 Arg Thr Gly Arg Gly Arg Arg Gly Ile Tyr Arg Phe Val Thr Pro Gly  
 770 775 780  
 Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val Leu Cys Glu Cys Tyr  
 785 790 795 800  
 Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro Ala Glu Thr Ser Val  
 805 810 815  
 Arg Leu Arg Ala Tyr Leu Asn Thr Pro Gly Leu Pro Val Cys Gln Asp  
 820 825 830  
 His Leu Glu Phe Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile Asp  
 835 840 845  
 Ala His Phe Leu Ser Gln Thr Lys Gln Ala Gly Asp Asn Phe Pro Tyr  
 850 855 860  
 Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Gln Ala Pro Pro  
 865 870 875 880  
 Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu Lys Pro Thr  
 885 890 895  
 Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ala Val Gln Asn  
 900 905 910  
 Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr Ile Met Ala Cys Met  
 915 920 925  
 Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp Val Leu Val Gly Gly  
 930 935 940  
 Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val Val  
 945 950 955 960

Ile Val Gly Arg Ile Ile Leu Ser Gly Arg Pro Ala Ile Val Pro Asp  
 965 970 975

Arg Glu Leu Leu Tyr Gln Glu Phe Asp Glu Met Glu Glu Cys Ala Ser  
 980 985 990

His Leu Pro Tyr Ile Glu Gln Gly Met Gln Leu Ala Glu Gln Phe Lys  
 995 1000 1005

Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu Ala  
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Ala Ala Pro Val Val Glu Ser Lys Trp Arg Ala Leu Glu Thr Phe Trp  
 1025 1030 1035 1040

Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly  
 1045 1050 1055

Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala Ser Leu Met Ala Phe  
 1060 1065 1070

Thr Ala Ser Ile Thr Ser Pro Leu Thr Thr Gln Ser Thr Leu Leu Phe  
 1075 1080 1085

Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser Ala  
 1090 1095 1100

Ala Ser Ala Phe Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly Ser  
 1095 1110 1115 1120

Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala  
 1125 1130 1135

Gly Val Ala Gly Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu Met  
 1140 1145 1150

Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro Ala Ile Leu Glu Glu  
 1155 1160 1165

Ala Ser Glu Asp Val Val Cys Cys Ser Met Ser Tyr Thr Trp Thr Gly  
 1170 1175 1180

Ala Leu Glu Leu Leu Leu Leu Leu Leu Gly Leu Arg Leu Gln Leu  
 1185 1190 1195 1200

Ser Leu Gly Ile Ile Pro Val Glu Glu Asn Pro Asp Phe Trp Asn  
 1205 1210 1215

Arg Glu Ala Ala Glu Ala Leu Gly Ala Ala Lys Lys Leu Gln Pro Ala  
 1220 1225 1230

Gln Thr Ala Ala Lys Asn Leu Ile Ile Phe Leu Gly Asp Gly Met Gly  
 1235 1240 1245

Val Ser Thr Val Thr Ala Ala Arg Ile Leu Lys Gly Gln Lys Lys Asp  
 1250 1255 1260

Lys Leu Gly Pro Glu Ile Pro Leu Ala Met Asp Arg Phe Pro Tyr Val  
 1265 1270 1275 1280

Ala Leu Ser Lys Thr Tyr Asn Val Asp Lys His Val Pro Asp Ser Gly  
 1285 1290 1295  
 Ala Thr Ala Thr Ala Tyr Leu Cys Gly Val Lys Gly Asn Phe Gln Thr  
 1300 1305 1310  
 Ile Gly Leu Ser Ala Ala Ala Arg Phe Asn Gln Cys Asn Thr Thr Arg  
 1315 1320 1325  
 Gly Asn Glu Val Ile Ser Val Met Asn Arg Ala Lys Lys Ala Gly Lys  
 1330 1335 1340  
 Ser Val Gly Val Val Thr Thr Arg Val Gln His Ala Ser Pro Ala  
 345 1350 1355 1360  
 Gly Thr Tyr Ala His Thr Val Asn Arg Asn Trp Tyr Ser Asp Ala Asp  
 1365 1370 1375  
 Val Pro Ala Ser Ala Arg Gln Glu Gly Cys Gln Asp Ile Ala Thr Gln  
 1380 1385 1390  
 Leu Ile Ser Asn Met Asp Ile Asp Val Ile Leu Gly Gly Arg Lys  
 1395 1400 1405  
 Tyr Met Phe Pro Met Gly Thr Pro Asp Pro Glu Tyr Pro Asp Asp Tyr  
 1410 1415 1420  
 Ser Gln Gly Gly Thr Arg Leu Asp Gly Lys Asn Leu Val Gln Glu Trp  
 425 1430 1435 1440  
 Leu Ala Lys Arg Gln Gly Ala Arg Tyr Val Trp Asn Arg Thr Glu Leu  
 1445 1450 1455  
 Met Gln Ala Ser Leu Asp Pro Ser Val Thr His Leu Met Gly Leu Phe  
 1460 1465 1470  
 Glu Pro Gly Asp Met Lys Tyr Glu Ile His Arg Asp Ser Thr Leu Asp  
 1475 1480 1485  
 Pro Ser Leu Met Glu Met Thr Glu Ala Ala Leu Arg Leu Leu Ser Arg  
 1490 1495 1500  
 Asn Pro Arg Gly Phe Phe Leu Phe Val Glu Gly Gly Arg Ile Asp His  
 505 1510 1515 1520  
 Gly His His Glu Ser Arg Ala Tyr Arg Ala Leu Thr Glu Thr Ile Met  
 1525 1530 1535  
 Phe Asp Asp Ala Ile Glu Arg Ala Gly Gln Leu Thr Ser Glu Glu Asp  
 1540 1545 1550  
 Thr Leu Ser Leu Val Thr Ala Asp His Ser His Val Phe Ser Phe Gly  
 1555 1560 1565  
 Gly Tyr Pro Leu Arg Gly Ser Cys Ile Phe Gly Leu Ala Pro Gly Lys  
 1570 1575 1580  
 Ala Arg Asp Arg Lys Ala Tyr Thr Val Leu Leu Tyr Gly Asn Gly Pro  
 585 1590 1595 1600

Gly Tyr Val Leu Lys Asp Gly Ala Arg Pro Asp Val Thr Glu Ser Glu  
 1605 1610 1615

Ser Gly Ser Pro Glu Tyr Arg Gln Gln Ser Ala Val Pro Leu Asp Glu  
 1620 1625 1630

Glu Thr His Ala Gly Glu Asp Val Ala Val Phe Ala Arg Gly Pro Gln  
 1635 1640 1645

Ala His Leu Val His Gly Val Gln Glu Gln Thr Phe Ile Ala His Val  
 1650 1655 1660

Met Ala Phe Ala Ala Cys Leu Glu Pro Tyr Thr Ala Cys Asp Leu Ala  
 665 1670 1675 1680

Pro Pro Ala Gly Thr Thr Asp Ala Ala His Pro Gly  
 1685 1690

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<211> 152

<212> PRT

<213> Artificial Sequence

<400> 19

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 1 5 10 15

Arg Lys Leu Ala Ser Arg Leu Met Pro Ser Glu Gln Trp Lys Gly Ile  
 20 25 30

Ile Ala Val Ser Arg Gly Leu Val Pro Gly Ala Leu Leu Ala Arg  
 35 40 45

Glu Leu Gly Ile Arg His Val Asp Thr Val Cys Ile Ser Ser Tyr Asp  
 50 55 60

His Asp Asn Gln Arg Glu Leu Lys Val Leu Lys Arg Ala Glu Gly Asp  
 65 70 75 80

Gly Glu Gly Phe Ile Val Ile Asp Asp Leu Val Asp Thr Gly Gly Thr  
 85 90 95

Ala Val Ala Ile Arg Glu Met Tyr Pro Lys Ala His Phe Val Thr Ile  
 100 105 110

Phe Ala Lys Pro Ala Gly Arg Pro Leu Val Asp Asp Tyr Val Val Asp  
 115 120 125

Ile Pro Gln Asp Thr Trp Ile Glu Gln Pro Trp Asp Met Gly Val Val  
 130 135 140

Phe Val Pro Pro Ile Ser Gly Arg  
 145 150

<210> 20

<211> 85

<212> PRT

<213> Artificial Sequence

<400> 20  
 Phe Cys Glu Arg Met Ala Asn Glu Gly Lys Ile Val Ile Val Ala Ala  
 1 5 10 15

Leu Asp Gly Thr Phe Gln Arg Lys Pro Phe Asn Asn Ile Leu Asn Leu  
 20 25 30

Ile Pro Leu Ser Glu Met Val Val Lys Leu Thr Ala Val Cys Met Lys  
 35 40 45

Cys Phe Lys Glu Ala Ser Phe Ser Lys Arg Leu Gly Glu Glu Thr Glu  
 50 55 60

Ile Glu Ile Ile Gly Gly Asn Asp Met Tyr Gln Ser Val Cys Arg Lys  
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Cys Tyr Ile Asp Ser  
 85

<210> 21

<211> 286

<212> PRT

<213> Artificial Sequence

<400> 21

Met Ser Ile Gln His Phe Arg Val Ala Leu Ile Pro Phe Phe Ala Ala  
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Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr Leu Val Lys Val Lys  
 20 25 30

Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu Leu Asp  
 35 40 45

Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg Pro Glu Glu Arg Phe  
 50 55 60

Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala Val Leu Ser  
 65 70 75 80

Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly Arg Arg Ile His Tyr Ser  
 85 90 95

Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His Leu Thr  
 100 105 110

Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala Ala Ile Thr Met Ser  
 115 120 125

Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr Ile Gly Gly Pro Lys  
 130 135 140

Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp His Val Thr Arg Leu  
 145 150 155 160

Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg  
 165 170 175

Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr Leu Arg Lys Leu Leu  
 180 185 190

Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp  
 195 200 205

Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu Arg Ser Ala Leu Pro  
 210 215 220

Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly Glu Arg Gly Ser  
 225 230 235 240

Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile  
 245 250 255

Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr Met Asp Glu Arg Asn  
 260 265 270

Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile Lys His Trp  
 275 280 285

<210> 22

<211> 220

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Sac 1/SEAP/Bam  
 H1 construct

<400> 22

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 atcccaaggatc aggaggagaa cccggacttc tggaaccgcg aggcagccga ggccctgggt 120  
 gccgccaaga agctgcagcc tgcacagaca gccgccaaga acctcatcat ctccctggc 180  
 gatggatgg gggtgtctac ggtgacagct gccaggatcc 220

<210> 23

<211> 88

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: amino acid  
 fragment of the HCV polyprotein

<400> 23

Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu Leu Ile Ala Gln Ala  
 1 5 10 15

Glu Ala Ala Leu Glu Asn Leu Val Val Leu Asn Ser Ala Ser Val Ala  
 20 25 30

Gly Ala His Gly Ile Leu Ser Phe Leu Val Phe Phe Cys Ala Ala Trp  
 35 40 45

Tyr Ile Lys Gly Arg Leu Val Pro Gly Ala Thr Tyr Ala Leu Tyr Gly  
 50 55 60

Val Trp Pro Leu Leu Leu Leu Ala Leu Pro Pro Arg Ala Tyr  
 65 70 75 80

Ala Met Asp Arg Glu Met Ala Ala  
 85

<210> 24  
<211> 260  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: DNA fragment  
 coding for an amino acid fragment of the HCV  
 polyprotein

<400> 24  
 gcacgtgtct gtgcctgctt gtggatgatg ctgctgatag cccaggccga ggccgccttg 60  
 gagaacctgg tggcctcaa tgcggcgctct gtggccggcg cacatggcat cctctcccttc 120  
 ctttgttct tctgtgccgc ctggatcacatc aaaggcaggc tggccctgg ggccgcataat 180  
 gctctttatg gcgtgtggcc gctgctcctg ctcttgctgg cattaccacc gcgagcttac 240  
 gccatggacc gggagatggc 260

<210> 25  
<211> 177  
<212> PRT  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: amino acid  
 fragment of the HCV polyprotein

<400> 25  
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 Gln Phe Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Thr Lys Gln  
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 Ala Glu Ala Ala Ala Pro Val Val Glu Ser Lys Trp Arg Ala Leu Glu  
 35 40 45  
 Thr Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr  
 50 55 60  
 Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala Ser Leu  
 65 70 75 80  
 Met Ala Phe Thr Ala Ser Ile Thr Ser Pro Leu Thr Thr Gln Ser Thr  
 85 90 95  
 Leu Leu Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu Ala Pro  
 100 105 110

Pro Ser Ala Ala Ser Ala Phe Val Gly Ala Gly Ile Ala Gly Ala Ala  
115 120 125

Val Gly Ser Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly  
130 135 140

Tyr Gly Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Val Met Ser  
145 150 155 160

Gly Glu Met Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro Ala Ile  
165 170 175

Leu

<210> 26

<211> 528

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA fragment  
coding for an amino acid fragment of the HCV  
polyprotein

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aaagcgctcg ggttactgca aacagccacc aaacaagcgg aggctgctgc tccctgttgt 120  
gagtccttggtggc tgagacattc tggcgaagc acatgtggaa ttcatcaggc 180  
gggatacagt acttagcagg cttatccact ctgcctggaa accccgcaat agcatcattg 240  
atggcattca cagcctctat caccagcccg ctcaccaccc aaagtaccct cctgtttAAC 300  
atcttggggg ggtgggtggc tgcccaactc gcccccccca gcgccgcttc ggctttcg 360  
ggcgccggca tcgcgggtgc ggctgttgtgc agcataggcc ttgggaagggt gcttggtggac 420  
attctggcgg gttatggagc aggagtggcc ggcgcgtcg tggccttAA ggtcatgagc 480  
ggcgagatgc cctccaccga ggacctggtc aatctacttc ctgccatc 528

<210> 27

<211> 33

<212> DNA

<213> primer

<400> 27

gcgcgcgaat tcatggcacg tgtctgtgcc tgc

33

<210> 28

<211> 33

<212> DNA

<213> primer

<400> 28

cgcgcgctcg aggatggcag gaagtagatt gac 33

<210> 29  
<211> 20  
<212> PRT  
<213> putative NS5A/5B cleavage site

<400> 29  
Glu Glu Ala Ser Glu Asp Val Val Cys Cys Ser Met Ser Tyr Thr Trp  
1 5 10 15

Thr Gly Ala Leu  
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<210> 30  
<211> 33  
<212> DNA  
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<400> 30  
gcgcgccctcg aggaagctag tgaggatgtc gtc 33

<210> 31  
<211> 36  
<212> DNA  
<213> primer

<400> 31  
cgcgcgaggc tccaaggcgc ctgtccatgt gtagga 36

<210> 32  
<211> 69  
<212> DNA  
<213> primer

<400> 32  
ctcgaggaag ctagtgagga tgtcgctgc tgctcaatgt cctacacatg gacaggcgcc 60  
ttggagctc 69

<210> 33  
<211> 6  
<212> PRT  
<213> HCV/SEAP 6 amino acid fragment

<400> 33  
Met Gly Ile Pro Gln Phe  
1 5

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/17440

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :Please See Extra Sheet.

US CL :435/5, 6, 23, 320.1; 530/350; 536/23.2

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/5, 6, 23, 320.1; 530/350; 536/23.2

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WEST/ALL; Dialog

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	HIROWATARI, Y. A Novel Method for Analysis of Viral Proteinase Activity Encoded by Hepatitis C Virus in Cultured Cells. Analytical Biochemistry. 1995, pages 113-120, see entire document.	1-41
Y	CHO, Y.-G. et al. In vivo assay for hepatitis C viral serine protease activity using a secreted protein. Journal of Virological Methods. 1998, Vol. 72, pages 109-115, see entire document.	1-41
Y	SONG, O-K. et al. Development of an in vivo Assay System Suitable for Screening Inhibitors of Hepatitis C Viral Protease. Molecular Cells. 1996, Vol. 6, No. 2, pages 183-189, see entire document.	1-41

 Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

22 NOVEMBER 1999

Date of mailing of the international search report

14 DEC 1999

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Facsimile No. (703) 305-3230

**INTERNATIONAL SEARCH REPORT**

International application No.

PCT/US99/17440

**C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,721,133 A (DASMAHAPATRA) 24 February 1998, see entire document.	1-41
A	US 5,739,002 A (DE FRANCESCO et al.) 14 April 1998.	1-41
A	INOUE, H. et al. Novel Assay System for Hepatitis C Virus Serine Protease Inhibitors. Antiviral Research. 1995, Vol. 26, No. 3. Abstract 122, page A289.	1-41

**INTERNATIONAL SEARCH REPORT**

International application No.

PCT/US99/17440

**A. CLASSIFICATION OF SUBJECT MATTER:**

IPC (6):

G01N 33/576; C12Q 1/68; G03C 5/00; C12N 15/51; C07K 14/18; C07H 21/04